

Filtering the NMR Spectra of Mixtures by Coordination to Paramagnetic  $\text{Cu}^{2+}$ 

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Cite This: *Anal. Chem.* 2022, 94, 10907–10911

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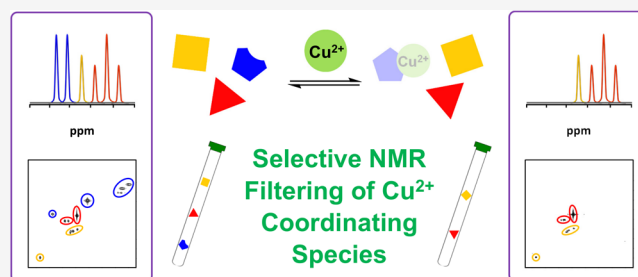


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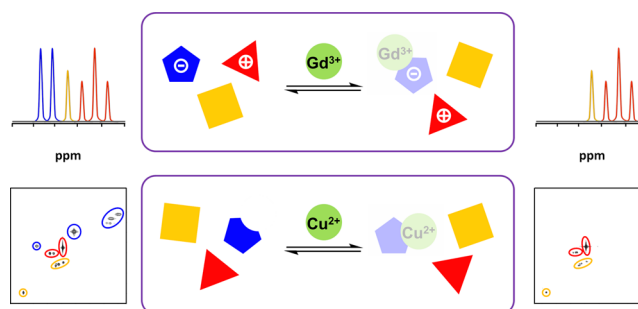


Supporting Information

**ABSTRACT:** The paramagnetic spin relaxation (PSR) filter allows the selective NMR signal suppression of components in mixtures according to their complexation ability to a paramagnetic ion. It relies on the faster relaxation of nuclei in paramagnetic environments and thus is complementary to classical diffusion and relaxation filters. So far, the PSR filter has established  $\text{Gd}^{3+}$  as the sole PSR agent, restricting the paramagnetic filtering repertoire. Herein, we present  $\text{Cu}^{2+}$  as a robust PSR agent with characteristic filtering properties. While  $\text{Gd}^{3+}$  depends on unspecific ion-pair interactions with anionic components,  $\text{Cu}^{2+}$  stands out for filtering species via ordered coordination complexes. An evaluation of the paramagnetic effect of  $\text{Cu}^{2+}$  over more than 50 small molecules and polymers has unveiled different sensitivities to  $\text{Cu}^{2+}$  (especially high for pyridines, diamines, polyamines, and amino alcohols) and precise filtering conditions for mixtures ( $^1\text{H}$ , COSY, and HMQC) that were challenged with a test bed of commercial drugs. The advantage of integrating  $\text{Cu}^{2+}$  and  $\text{Gd}^{3+}$  for the stepwise PSR filtering of complex mixtures is also shown.



As nature seldom provides pure compounds, chemists have devoted great efforts to the separation of complex mixtures. Fortunately, the NMR analysis of mixtures sidesteps the necessity of physical separations under certain conditions. NMR filters take advantage of differences in the diffusion coefficients and relaxation times of the components for the selective signal suppression of small molecules and macromolecules, respectively.<sup>1,2</sup> With the aim of widening the NMR filtering portfolio beyond molecular weight limits, our group has described the paramagnetic spin relaxation (PSR) filter,<sup>3,4</sup> which relies on the faster relaxation of nuclei in paramagnetic environments.<sup>5,6</sup> The addition of minute concentrations of  $\text{Gd}^{3+}$  (paramagnetic) to mixtures allows the selective suppression of particular components from the 1D and 2D spectra, according to their  $\text{Gd}^{3+}$  complexation ability (Figure 1). Since  $\text{Gd}^{3+}$  has the largest spin moment ( $S = 7/2$ ) and a high electronic correlation time ( $\tau_e$ , ca.  $10^{-8}$  s), the Solomon–Bloembergen–Morgan equations predict a selective decrease of transverse relaxation times ( $T_2$ ) for species in fast chemical exchange with  $\text{Gd}^{3+}$ .<sup>7–9</sup> The inverse proportionality between  $T_2$  and the spectral line width<sup>10</sup> leads to their selective signal suppression, without affecting the resolution and chemical shift of other components in the mixture. Since complexation to  $\text{Gd}^{3+}$  is mainly electrostatic (ion-pair), the PSR filter affects more anionic species than neutral and cationic species. Successful applications of the PSR filter include the sequential NMR filtering of mixtures of interest in the pharmaceutical and food industries<sup>4,11</sup> and the fast screening of DNA ligands.<sup>12</sup> Interestingly, the PSR filter benefits from a rather low



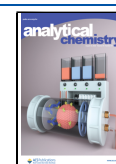
**Figure 1.** PSR filters based on  $\text{Gd}^{3+}$  (ion-pair) and  $\text{Cu}^{2+}$  (coordination) complexes.

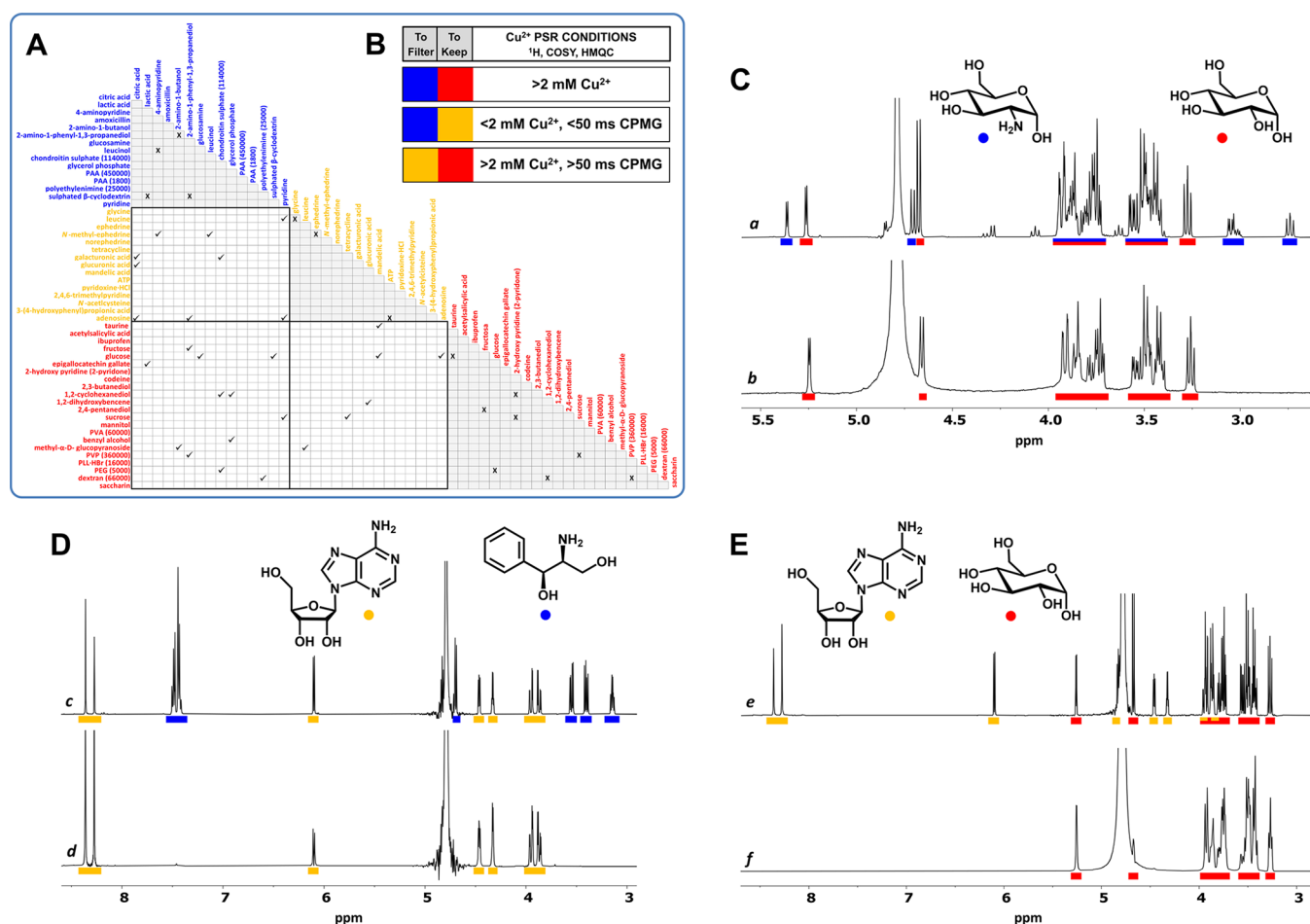
sensitivity to size and molecular weight that makes it compatible with classical relaxation and diffusion filters. The advantage of this is taken for the filtering of lower PSR-sensitive species, which leads to line-broadening rather than a full signal embedment in the baseline, by implementing a complementary short  $T_2$ -filter, such as the Carr–Purcell–Meiboom–Gill (CPMG).<sup>13,14</sup>

Received: May 6, 2022

Accepted: July 25, 2022

Published: July 27, 2022





**Figure 2.** (A) Successful and failed PSR suppressions in two-component mixtures (D<sub>2</sub>O, 500 MHz). (B) PSR filtering conditions for selective <sup>1</sup>H, COSY, and HMQC suppressions. Representative examples of selective suppressions between Blue-Yellow-Red categories. <sup>1</sup>H NMR spectra (D<sub>2</sub>O, 500 MHz, 300 K) of a mixture of the following: (C) glucosamine (2 mg/mL) and glucose (2 mg/mL) before (a) and after (b) the addition of Cu<sup>2+</sup> (2 mM), (D) 2-amino-1-phenyl-1,3-propanediol (1 mg/mL) and adenosine (3 mg/mL) before (c) and after (d) the addition of Cu<sup>2+</sup> (0.16 mM) + T<sub>2</sub>-filter (CPMG, 30 ms), and (E) adenosine (1.2 mg/mL) and glucose (1.6 mg/mL) before (e) and after (f) the addition of Cu<sup>2+</sup> (13 mM) + T<sub>2</sub>-filter (CPMG, 100 ms).

While the PSR filter based on ion-pair Gd<sup>3+</sup>-complexes is highly efficient in simplifying the NMR analysis of complex mixtures, the development of PSR filters with other paramagnetic ions displaying alternative complexation modes would greatly extend the utility of this technology. Herein, we describe our efforts toward a PSR filter based on the more coordinating Cu<sup>2+</sup> ( $S = 1/2$ ,  $\tau_s$  ca.  $10^{-9}$  s), a paramagnetic ion aimed at selectively filtering the NMR signals of species via coordination complexes (Figure 1).

The feasibility of a Cu<sup>2+</sup> PSR filter was confirmed by the selective suppression of glucosamine in the presence of glucose (Figure 2C), a suppression unfeasible to reproduce in the presence of Gd<sup>3+</sup> (Figure S4) because of the similar sensitivity of both components to this ion.<sup>4</sup> Conversely, the coordinating 1,2-amino alcohol moiety of glucosamine ensures a selective filtering in the presence of Cu<sup>2+</sup>.

The scope of Cu<sup>2+</sup> as coordinating PSR agent was assessed by analyzing the paramagnetic effect on the <sup>1</sup>H NMR spectra of a collection of more than 50 small molecules and polymers of interest in the pharmaceutical and food industries, which display a large variety of functional groups. Depending on the extent of signal broadening (from no effect to complete suppression), these species were assigned to seven groups

(Table 1), with the more sensitive ones comprising highly coordinating compounds. <sup>1</sup>H NMR spectra of representative molecules in the upper, medium, and bottom parts of Table 1, recorded in the absence/presence of Cu<sup>2+</sup>, are shown in Figures S1–S3: (1*S*,2*S*)-2-amino-1-phenyl-1,3-propanediol, adenosine, sucrose. As a rule of thumb, the broadening effect of Cu<sup>2+</sup> on the <sup>1</sup>H NMR spectra of pyridines, diamines, polyamines, and amino alcohols was considerably higher than with Gd<sup>3+</sup>,<sup>4</sup> in consistency with a higher coordination ability.

Next, the possibility of performing selective suppressions by Cu<sup>2+</sup> among the seven groups in Table 1 was assessed in two-component mixtures (Figure 2A). It was confirmed that the more distant the groups, the easier the selective suppressions. Also, the impossibility of performing suppressions within a single group and between some neighboring groups. As a result, the initial groups (reflecting the paramagnetic broadening effect) were reduced to just three categories (designated as Blue, Yellow, and Red), according to their ease of suppression by Cu<sup>2+</sup>. In addition, from data in Figure 2A, general conditions for selective <sup>1</sup>H, COSY, and HMQC suppressions between categories were determined (concentration of Cu<sup>2+</sup>/length of a complementary CPMG T<sub>2</sub>-filter; Figure 2B):

**Table 1. Paramagnetic Broadening Effect (Groups 1–7) and Ease of Suppression by Cu<sup>2+</sup> (Blue, Yellow, Red Categories) in <sup>1</sup>H NMR (4 mg/mL in D<sub>2</sub>O, 500 MHz, 300 K)<sup>a</sup>**

	Compound	Type	Compound	Type
7	citric acid	hydroxy acid	<i>N</i> -acetylcysteine	amino acid
	lactic acid	hydroxy acid	3-(4-hydroxyphenyl)	hydroxy acid
	4-aminopyridine	pyridine	propionic acid	hydroxy acid
	amoxicillin	amino acid	adenosine	nucleoside
6	2-amino-1-butanol	amino alcohol	taurine	amino sulfonic acid
	2-amino-1-phenyl-1,3-propanediol	amino alcohol	acetylsalicylic acid	carboxylic acid
	glucosamine	amino alcohol	ibuprofen	carboxylic acid
	leucine	amino alcohol	fructose	monosaccharide
5	chondroitin sulphate	glycosaminoglycan	glucose	monosaccharide
	glycerol phosphate	phosphate	epigallocatechin gallate	polyphenol
	PAA (450000)	polyacid	2-hydroxypyridine (2-pyridone)	pyridone
	PAA (1800)	polyacid	codeine	amino alcohol
4	polyethylenimine	polyamine	2,3-butanediol	diol
	sulphated β-cyclodextrin	polysulphate	1,2-cyclohexanediol	diol
	pyridine	pyridine	1,2-dihydroxybenzene	diol
	glycine	amino acid	2,4-pentanediol	diol
3	leucine	amino acid	sucrose	disaccharide
	ephedrine	amino alcohol	mannitol	polyol
	<i>N</i> -methylephedrine	amino alcohol	PVA	polyol
	norephedrine	amino alcohol	benzyl alcohol	alcohol
2	tetracycline	amine, polyol	methyl-α-D-glucopyranoside	monosaccharide
	galacturonic acid	hydroxy acid	PVP	polyamide
	glucuronic acid	hydroxyl acid	PLL-HBr	polyamine
	mandelic acid	hydroxy acid	PEG	polyether
1	ATP	nucleotide	dextran	polysaccharide
	pyridoxine-HCl	pyridine	saccharin	sulfonamide
	2,4,6-trimethylpyridine	pyridine		

<sup>a</sup>Group 7: suppression of all signals at <1 mM Cu<sup>2+</sup>, Group 6: suppression of all signals at >1 mM Cu<sup>2+</sup>, Group 5: broadening of all signals at >1 mM Cu<sup>2+</sup>, Group 4: suppression of some signals at >1 mM Cu<sup>2+</sup>, Group 3: broadening of some signals at >1 mM Cu<sup>2+</sup>, Group 2: reduced signal resolution at >1 mM Cu<sup>2+</sup>, Group 1: no effect on signal resolution at >1 mM Cu<sup>2+</sup>.

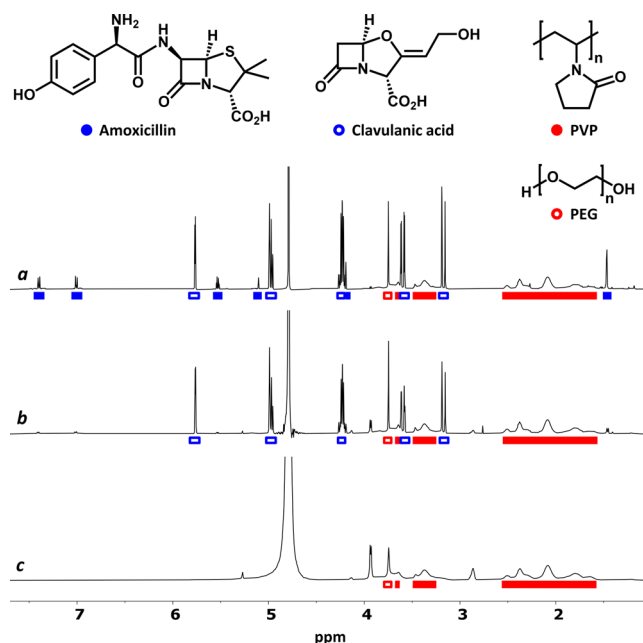
- Blue-Red: > 2.0 mM Cu<sup>2+</sup>
- Blue-Yellow: < 2.0 mM Cu<sup>2+</sup>, < 50 ms CPMG
- Yellow-Red: > 2.0 mM Cu<sup>2+</sup>, > 50 ms CPMG

Figure 2 depicts representative examples of successful suppressions within these three categories. For instance, the aforementioned glucosamine/glucose suppression can be easily rationalized now, considering their respective inclusion into the Blue and Red categories (Figure 2C). Similarly, Figure 2D,E show selective suppressions that exploit the use of complementary *T*<sub>2</sub>-filters between components of contiguous categories. Remarkably, none of these suppressions could be realized with Gd<sup>3+</sup> as PSR ion (Figures S4–S6), confirming the coordination ability of Cu<sup>2+</sup> as responsible of the selectivity achieved. Other representative spectra of successful (different categories) and unfruitful (same category) suppressions in the two-component mixtures shown in Figure 2A are included in the SI (Figures S7–S12). Additional advantages that emerged on the use of Cu<sup>2+</sup> vs Gd<sup>3+</sup> as PSR agent include the possibility of working in a wider pH-range (Gd<sup>3+</sup> tends to precipitate at pH > 7.0) and a smaller broadening effect over the residual HOD signal.

The reliability of Table 1 to predict Cu<sup>2+</sup> PSR filters in complex mixtures was challenged with a test bed of commercial drugs, namely (i) an amoxicillin/clavulanic acid antibiotic (amoxicillin, clavulanic acid, PEG, PVP); (ii) Cariban, a drug to treat nausea and vomiting in pregnancy (doxylamine, pyridoxine, sucrose); (iii) the antibiotic Proderma (doxycycline, sucrose); and (iv) Atepodin, a medicine for the treatment and diagnosis of supraventricular tachycardia (adenosine triphosphate, glycine, benzyl alcohol).

We started analyzing Amoxicillin/Clavulanic acid Cinfamed, an antibiotic that contains two active ingredients, the penicillin-like antibiotic amoxicillin (Blue) and the beta-lactamase inhibitor clavulanic acid (not included in Table 1 but expected to be Blue by similarity). NMR-visible excipients

comprise poly(ethylene glycol) (PEG) and polyvinylpyrrolidone (PVP), both Red species. Attending to the Blue and Red categories of the constituents, a selective PSR suppression was anticipated for amoxicillin and clavulanic acid in the presence of Cu<sup>2+</sup>. Figure 3 shows their selective filtering, confirming the

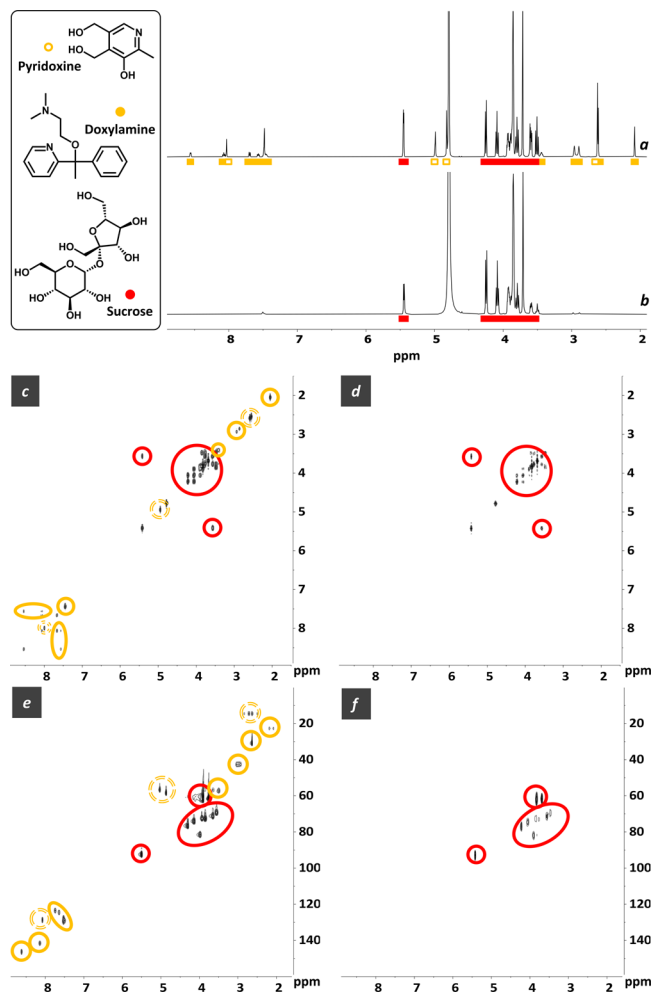


**Figure 3.** <sup>1</sup>H NMR spectra (D<sub>2</sub>O, 500 MHz, 300 K) of Amoxicillin/Clavulanic acid Cinfamed (10 mg/mL) before (a) and after the addition of 4 and 17 mM Cu<sup>2+</sup> (b and c, respectively).

predictive character of Table 1, and the three color categories proposed. When lower concentrations of Cu<sup>2+</sup> were assessed, it was even possible to attain a stepwise suppression of the Blue components, filtering first the more coordinating amoxicillin. Interestingly, the use of Gd<sup>3+</sup> did not afford a clean suppression of any component, confirming Cu<sup>2+</sup> as a PSR agent with characteristic filtering properties.

Then, we proceeded to analyze three commercial drugs (Cariban, Proderma, Atepodin) containing Yellow and Red components, where a complementary CPMG filter was expected for successful suppressions. Cariban is a medicine used to treat nausea and vomiting in pregnant women that contains doxylamine (antihistamine) and pyridoxine (vitamin B<sub>6</sub>) as active ingredients, both substituted pyridines that belong to the Yellow category. The mixture also includes sucrose (Red) as NMR-visible excipient. As predicted, the only addition of Cu<sup>2+</sup> did not result in a clean filtering of doxylamine and pyridoxine. However, concomitant application of a short CPMG filter afforded their clean suppression, leaving unaffected the resolution and chemical shift of the sucrose signals (Figure 4). Remarkably, the fidelity of the PSR-CPMG filter was also demonstrated in 2D COSY and HMQC experiments, where the CPMG sequence was used as an excitation block replacing the first excitation pulse.<sup>15</sup> Not unexpectedly, the use of Gd<sup>3+</sup> as PSR agent was again unsuccessful, either in the absence or presence of CPMG filters.

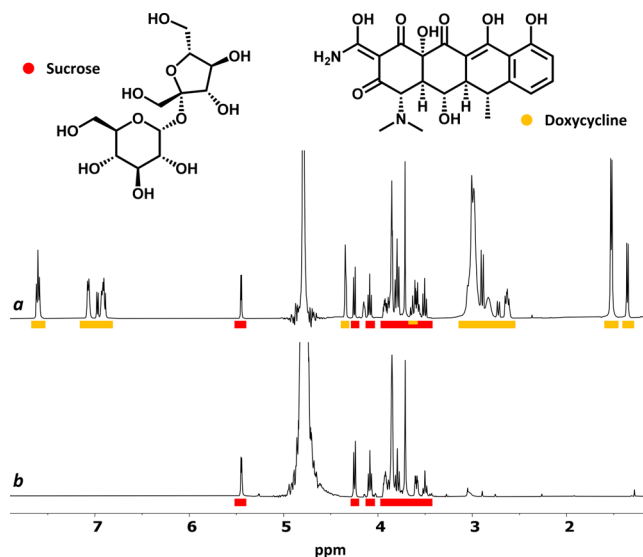
The antibiotic Proderma contains two NMR-visible components, the active ingredient doxycycline (Yellow) and sucrose (Red) as excipient. Here again, the direct filtering of the most sensitive Yellow component was unfeasible by the



**Figure 4.**  $^1\text{H}$ ,  $^1\text{H}-^1\text{H}$  COSY, and  $^1\text{H}-^{13}\text{C}$  HMQC spectra ( $\text{D}_2\text{O}$ , 500 MHz, 300 K) of Cariban (72 mg/mL) before (a, c, e) and after (b, d, f) the addition of  $\text{Cu}^{2+}$  (13 mM) +  $T_2$ -filter (CPMG, 75 ms).

sole addition of  $\text{Cu}^{2+}$ . However, implementation of a simultaneous short CPMG filter allowed the clean suppression of doxycycline (Figure 5). Very similar filtering conditions also allowed the efficient filtering of the Yellow components (adenosine triphosphate and glycine) of Atepodin (Figure S13).

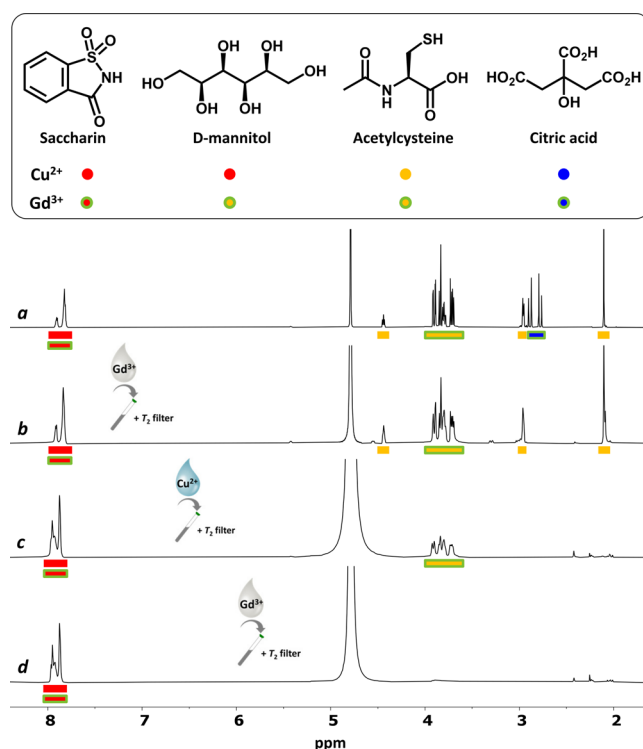
Having established the utility of  $\text{Cu}^{2+}$  as PSR agent with filtering properties dependent on the coordination ability of the components in a mixture (rather than ion-pair interactions for  $\text{Gd}^{3+}$ ), we decided to assess the integration of both paramagnetic ions in the filtering of complex mixtures. To this end, we selected Acetilcisteina Mylan, a commercial mucolytic drug composed of acetylcysteine ( $\text{Cu}$ Yellow/ $\text{Gd}$ Yellow) as active ingredient, citric acid ( $\text{Cu}$ Blue/ $\text{Gd}$ Blue) as excipient, and two sweeteners, D-mannitol ( $\text{Cu}$ Red/ $\text{Gd}$ Yellow) and sodium saccharin ( $\text{Cu}$ Red/ $\text{Gd}$ Red). Attending to the Blue-Yellow-Red category of the components toward  $\text{Gd}^{3+}$ , we have previously reported the sequential suppression of citric acid in a first step, followed by the simultaneous suppression of acetylcysteine and D-mannitol (both  $\text{Gd}$ Yellow components).<sup>4</sup> Herein, the stronger  $\text{Cu}^{2+}$ -coordination of acetylcysteine than D-mannitol (Table 1) has been exploited for the stepwise suppression of the three components in a way unattainable with a single PSR agent. Thus, as shown in Figure 6, after an initial suppression



**Figure 5.**  $^1\text{H}$  NMR spectra ( $\text{D}_2\text{O}$ , 500 MHz, 300 K) of Proderma (4 mg/mL) before (a) and after (b) the addition of  $\text{Cu}^{2+}$  (4 mM) +  $T_2$ -filter (CPMG, 75 ms).

of citric acid with  $\text{Gd}^{3+}$ , acetylcysteine was filtered with  $\text{Cu}^{2+}$ , followed by a final suppression of D-mannitol with  $\text{Gd}^{3+}$ .

Application of the PSR filter with  $\text{Cu}^{2+}$  starts with the assignment of the individual components in a mixture to the Blue-Yellow-Red categories. As a first approach, users are



**Figure 6.** NMR-visible components of Acetilcisteina Mylan and their respective Blue-Yellow-Red categories toward  $\text{Cu}^{2+}$  and  $\text{Gd}^{3+}$ .  $^1\text{H}$  NMR spectra ( $\text{D}_2\text{O}$ , 500 MHz, 300 K) of Acetilcisteina Mylan (25 mg/mL) supplemented with saccharin (12.5 mg/mL) before (a) and after the addition of (b)  $\text{Gd}^{3+}$  (25  $\mu\text{M}$ ) +  $T_2$ -filter (CPMG, 25 ms), (c)  $\text{Cu}^{2+}$  (2.5 mM) +  $T_2$ -filter (CPMG, 250 ms), and (d)  $\text{Gd}^{3+}$  (0.2 mM) +  $T_2$ -filter (CPMG, 300 ms).

advised to find structural similarities between the species in a mixture of interest and those in Table 1. Nevertheless, for a proper inclusion of species in the Blue-Yellow-Red categories, the paramagnetic effect of  $\text{Cu}^{2+}$  on their  $^1\text{H}$  NMR spectra should be determined as described in Table 1 (extent of signal broadening as a function of the concentration of  $\text{Cu}^{2+}$ ). Once the species have been assigned to the three categories, selective suppressions could be expected by application of the PSR conditions shown in Figure 2B. While clean suppressions operate for Blue species in the presence of Red ones by the simple addition of mM concentrations of  $\text{Cu}^{2+}$ , the selective filtering of species from contiguous categories (Blue-Yellow and Yellow-Red) are unfeasible by the sole addition of  $\text{Cu}^{2+}$ , being necessary the implementation of simultaneous CPMG filters. Although the selective filtering of species within a category might work in specific examples using increasing concentrations of  $\text{Cu}^{2+}$  or tuning the length of CPMG filters, this will not be of general application because the  $^1\text{H}$   $T_2$  values of the species in a category will level down in the presence of a PSR agent, making unlikely their selective suppression.

In conclusion,  $\text{Cu}^{2+}$  is presented as a robust PSR agent with characteristic NMR filtering properties different than  $\text{Gd}^{3+}$ , the archetypal PSR agent so far. Not only do the paramagnetic properties change between nuclei, but also their complexation modes differ, offering the opportunity to tune the outcome of the PSR filter. While  $\text{Gd}^{3+}$  relies on the ion-pair complexation ability of the components in a mixture (mainly anionic species),  $\text{Cu}^{2+}$  stands out because of a greater capacity of filtering species that participate in coordination complexes, such as pyridines, diamines, polyamines, and amino alcohols. An evaluation of the paramagnetic effect of  $\text{Cu}^{2+}$  over more than 50 small molecules and polymers has unveiled three categories of compounds (Blue-Yellow-Red categories according to their ease of suppression by  $\text{Cu}^{2+}$ ) and precise filtering conditions for  $^1\text{H}$ , COSY, and HMQC between them. The integration of the specific filtering properties of  $\text{Cu}^{2+}$  and  $\text{Gd}^{3+}$  as PSR agents to the analysis of complex mixtures has been also demonstrated, widening the horizons of the PSR technology to quality control, natural product extracts, or the metabolic profiling of biological samples. Finally, having demonstrated the utility of  $\text{Cu}^{2+}$  as PSR agent, a more precise assignment of species to the Blue-Yellow-Red categories is envisaged using the transverse relaxation enhancement ( $R_{2p}$ ), as previously done for  $\text{Gd}^{3+}$ .<sup>4</sup> This approach, which involves the analysis of the  $^1\text{H}$   $T_2$  values of the species of interest in the absence and presence of  $\text{Cu}^{2+}$ , will be the focus of our investigations in the future and reported in due time.

## ■ ASSOCIATED CONTENT

### SI Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.analchem.2c01983>.

Materials, methods, and NMR spectra (PDF)

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## Notes

The authors declare no competing financial interest.

## ■ ACKNOWLEDGMENTS

This work was supported by the Spanish Ministry of Science and Innovation (RTI2018-102212-B-I00 and PID2021-127684OB-I00), the Xunta de Galicia (ED431C 2018/30, and Centro singular de investigación de Galicia accreditation 2019–2022, ED431G2019/03), Axencia Galega de Innovación (IN845D 2020/09), and the European Union (European Regional Development Fund-ERDF).

## ■ REFERENCES

- (1) Novoa-Carballal, R.; Fernandez-Megia, E.; Jimenez, C.; Riguera, R. *Nat. Prod. Rep.* **2011**, *28*, 78–98.
- (2) Bingol, K.; Brusweiler-Li, L.; Li, D.; Zhang, B.; Xie, M.; Bruschweiler, R. *Bioanalysis* **2016**, *8*, 557–573.
- (3) Fernandez-Megia, E.; Correa, J.; Novoa-Carballal, R.; Riguera, R. *J. Am. Chem. Soc.* **2007**, *129*, 15164–15173.
- (4) Correa, J.; Pinto, L. F.; Riguera, R.; Fernandez-Megia, E. *Anal. Chem.* **2015**, *87*, 760–767.
- (5) Bertini, I.; Luchinat, C.; Aime, S. *Coord. Chem. Rev.* **1996**, *150*, 77–110.
- (6) Helm, L. *Prog. Nucl. Magn. Reson. Spectrosc.* **2006**, *49*, 45–64.
- (7) Kowalewski, J.; Nordenskiöld, L.; Benetis, N.; Westlund, P.-O. *Prog. Nucl. Magn. Reson. Spectrosc.* **1985**, *17*, 141–185.
- (8) Solomon, I. *Phys. Rev.* **1955**, *99*, 559–565.
- (9) Bloembergen, N. *J. Chem. Phys.* **1957**, *27*, 572–573.
- (10) La Mar, G. N.; de Ropp, J. S. In *Biological Magnetic Resonance*; Berliner, L. J., Reuben, J., Eds.; Plenum Press: New York, 1993; Vol. 12, pp 1–78.
- (11) Correa, J.; Pinto, L. F.; Zhao, L.; Riguera, R.; Fernandez-Megia, E. *Chem. Eur. J.* **2018**, *24*, 19236–19242.
- (12) Pinto, L. F.; Correa, J.; Zhao, L.; Riguera, R.; Fernandez-Megia, E. *ACS Omega* **2018**, *3*, 2974–2983.
- (13) Meiboom, S.; Gill, D. *Rev. Sci. Instrum.* **1958**, *29*, 688–691.
- (14) Carr, H. Y.; Purcell, E. M. *Phys. Rev.* **1954**, *94*, 630–638.
- (15) Williams, P. G.; Saunders, J. K.; Dyne, M.; Mountford, C. E.; Holmes, K. T. *Magn. Reson. Med.* **1988**, *7*, 463–471.