

1 **Can proteomics contribute to biomonitoring of aquatic**
2 **pollution? A critical review**
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13 **Abstract**

14 Aquatic pollution is one of the greatest environmental problems, and therefore its control
15 represents one of the major challenges in this century. In recent years, proteomics has
16 emerged as a powerful tool for searching protein biomarkers in the field of pollution
17 biomonitoring. For biomonitoring marine contamination, there is a consensus that bivalves
18 are preferred organisms to assess organic and inorganic pollutants. Thus, the bivalve
19 proteome was intensively studied, particularly the mussel. It is well documented that heavy
20 metal pollution and organic chemicals altered the structural proteins causing degradation of
21 tissues of molluscs. Also, it is well known that proteins involved in stress oxidative such as
22 glutathione and enzymes as catalase, superoxide dismutase or peroxisomes are overexpressed
23 in response to contaminants. Additionally, using bivalves, other groups of proteins proposed
24 as pollution biomarkers are the metabolic proteins. Even though other marine species are
25 used to monitor the pollution, the presence of proteomic tools in these studies is scarce.
26 Concerning freshwater pollution field, a great variety of animal species (fish and crustaceans)
27 are used as biomonitors in proteomics studies compared to plants that are scarcely analysed.
28 In fish species, proteins involved in stress oxidative such as heat shock family or proteins
29 from lipid and carbohydrate metabolism were proposed as candidate biomarkers. On the
30 contrary, for crustaceans there is a lack of proteomic studies individually assessing the
31 contaminants. Novel scenarios, including emerging contaminants and new threats, will
32 require proteomic technology for a systematic search of protein biomarkers and a greater
33 knowledge at molecular level of those cellular pathways induced by contamination.

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36 **Abbreviations**

37 Two-dimensional gel electrophoresis (2-DE)

38 Isobaric tags for relative and absolute quantitation (iTRAQ)

39 Mass spectrometry (MS)

40 Two-dimensional difference gel electrophoresis (DIGE)

41 Matrix-assisted laser desorption ionisation (MALDI)

42 Time-of-flight mass spectrometry (TOF)

43 Sequential window acquisition of all theoretical fragment ion spectra mass spectrometry
44 (SWATH-MS)

45 Liquid chromatography mass spectrometry (LC/MS)

46 Data-independent acquisition (DIA) strategies

47 Data-dependent acquisition (DDA) strategies

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49 1. Introduction

50 Most human activities have a negative impact on aquatic ecosystems causing a serious
51 environmental problem (EEA, 2018). Emissions and waste from industries, unregulated
52 household waste, mining, aquaculture, pesticides and harmful chemicals in agriculture and
53 livestock end up polluting marine and freshwater ecosystems around the world. Thus, large
54 quantities of pollutants are accumulated in coastal ecosystems (Islam and Tanaka, 2004;
55 Halpern et al., 2008), lakes, rivers, aquifers, and groundwater (Vörösmarty et al., 2010;
56 Behmel et al., 2016) posing a great threat to life in these environments. Therefore, water
57 quality control and management are an imperative issue for the health of ecosystems and
58 human populations (UNESCO, 2009).

59 Biomonitoring is an ecological, simple, and economical tool which can be used to
60 carry out the pollution control on aquatic ecosystems. **Biomonitoring is the assessment of a**
61 **chemical toxic into a living organism at level of molecules, organelles, cells, tissues, organs,**
62 **organisms, populations, communities, ecosystems and biomes. Quantitative variables related**
63 **to chemical, biochemical, morphological, physiological, behavioural or abundant**
64 **concentrations indicate the pollutant toxicity.** They seem to indicate certain characteristics of
65 the environment related to the level of pollution in the environment (Bargagli, 1998; Markert
66 et al., 2003). Thus, there are hundreds of studies on biomonitoring of water quality, most of
67 them focus on the bioconcentration or bioaccumulation of pollutants in organisms from
68 marine or freshwater environments (*e.g.* Debén et al., 2015; García-Seoane et al., 2018), as
69 well as those focus on studies at the community level (*e.g.* Pandey et al., 2018). However,
70 there are few studies at the molecular level that use biochemical biomarkers, such as proteins,
71 to biomonitor water pollution.

72 The final concentration of pollutant in the aquatic organisms is the result of the
73 balance between the concentration of the chemical substance in the organism and the water.
74 **However, studies on the bioaccumulation of pollutants in aquatic organisms have several**

75 limitations such as kinetic balance that have not yet been solved as reviewed Schöne and
76 Krause Jr (2016). For instance, *Mytilus trossulus* and *Mytilus californianus* exposure to low levels
77 of Cd resulting high kinetic differences in metal uptake and release (Lares and Orians, 1997).
78 Additionally, the direct bioaccumulation of the pollutants in the organisms, does not enough
79 information over toxic effects in organisms (Wang, 2016). On the other hand, biochemical
80 alterations would be useful because they can provide information on the toxic effects of
81 pollutants. Many protein biomarkers can be developed to detect and quantify a specific
82 pollutant. Indeed, it is known that several proteins act as biomarkers in a particular aquatic
83 organism in response to a chemical as discussed in this review. As we mentioned earlier,
84 there are few studies on biochemical biomarkers in aquatic pollution compared to the high
85 number on bioconcentration or bioaccumulation ones. This may be because there are a high
86 variability of biochemical alterations depending on the type of organism and pollutant being
87 studied. In addition, they are rarely used as complementary techniques. Studies using
88 biomarkers provide the concentrations of pollutants in the environment but not the
89 bioconcentrations in the organisms used. Therefore, the presence of pollutants in the
90 environment and biological responses should be studied thoroughly to expand the range of
91 protein biomarkers as well as search new biomonitoring organisms for which protein
92 biomarkers have been not identified. Integrating all these data could help us to stablish the
93 toxic mechanisms in the organisms.

94 Proteomics is a useful and effective tool for searching protein biomarkers in the field
95 of pollution biomonitoring. Proteomics and other OMICs techniques have the potential to
96 identify protein biomarkers as well as protein patterns which would become more robust
97 and specific markers of a particular pollutants (Armengaud, 2016). A proteomic approach
98 could help us to develop a broader view of the protein alterations, identifying the biomarker
99 candidates. Proteome analysis could provide an overall picture of structural and functional
100 cellular information and the response mechanism against aquatic pollution. These proteins

101 biomarkers after validation process, are useful as alternative biomarkers for routine
102 application (Schmit et al., 2018). All this information would enable to develop commercial
103 kits based on protein markers. In recent years, there are two main proteomic approaches
104 established: protein separation in gel and mass spectrometry analysis (Tholey and Becker,
105 2017). Electrophoretic and chromatographic techniques based on prior protein digestion are
106 called bottom-up approach. The other strategy, top-down, is based on analysis of intact
107 protein by means of separation and mass spectrometry. Specifically, the second choice in
108 combination with small protein biomarkers (below 30 kDa) could be easily and routinely
109 implemented to identify and quantify protein biomarkers (Tholey and Becker, 2017).
110 Additionally, the intense analysis of proteins would contribute to the identification of more
111 specific and sensitive markers of pollution. These novel tools about the knowledge of
112 complete proteomes **open up** new avenues for environmental sciences. Currently,
113 environmental proteomics is being developed as a key branch of environmental sciences,
114 providing more information about molecular responses to contaminants in living organisms
115 (Li et al., 2018).

116 In the present review, the recent proteomic literature in the field of water pollution
117 using biomonitors was provided. The aim was to identify the main aquatic species and the
118 proteomic techniques used to search new protein biomarkers as well as to propose new
119 strategies. A discussion about the constraints of several species due to limitations in databases
120 or technical problems was conducted in the following sections.

121 **2. Search and selection of literature**

122 A systematic search of literature was performed using different tools, webs and databases:
123 Google Scholar, Pubmed.gov (NCBI), Scopus and Web of Science (Clarivate Analytics). The
124 keywords used for the search were: “contamination”, “pollution”, “biomonitors”,
125 “bioindicators”, “aquatic”, “animals” and “plants” combined with “biomarker”,
126 “proteome”, “proteomics” and “protein biomarker”. Only original papers published from

127 2010 to 2020 were considered and all were carefully checked. The most appealing articles
128 were included in Tables 1 and 2 which yielded a total of 30 and 19 articles for marine and
129 freshwater environments, respectively.

130 **3. Biomonitoring of marine pollution**

131 The most abundant candidate species for biomonitoring marine pollution are invertebrates
132 which constitute a huge group of macroscopic species, highlighting among them the bivalve
133 molluscs for their key role in the ecosystems and for their great value to aquaculture industry
134 (Campos et al., 2012). Bivalves are used for biomonitoring due to ubiquitous distribution,
135 easy accessibility, high filtering capacity as well as high resistance to a wide range of pollutants
136 (Suárez-Ulloa et al., 2013). Even emerging pollutants such as microplastics (Ward et al.,
137 2019), nanoparticles (Zha et al., 2019) and other organic pollutants (Rodil et al., 2019) have
138 been studied on bivalves.

139 Indeed, mussels are widely used to monitor the coastal water pollution (*e.g.* heavy metals,
140 polycyclic aromatic hydrocarbons, polychlorinated biphenyls, etc.) as reviewed Beyer et al.
141 (2017). Accordingly, mussel watch programs define different environmental quality
142 parameters. For this purpose, mussels distributed stations throughout different locations and
143 for fixed periods of time were analysed (Jernelov, 1996). Currently, the scientific community
144 is developing an intensive research in order to standardize protocols for monitor of pollution
145 using mussels even in emergent pollutants (Li et al., 2019). In addition, there is an increasing
146 concern because bivalves as edible species can be a health risk for humans if the metal
147 pollutants concentration, such as Cd, Pb, Hg and As is too high (Ruiz-Fernández et al., 2018).
148 Indeed, there are several guidelines of different countries and associations with
149 recommendations of these trace elements (Stanković et al., 2011). For instance, for Cd
150 contamination, the salinity on the bioaccumulation and depuration are important factors to
151 safe oyster *Crassostrea gigas* consumption (Sun et al., 2018). In contrast, little is known about
152 effects of emerging pollutants and no maximum residue levels have yet been set. Few effects

153 of these contaminants are known. For example, microplastics that could cause negative
154 effects on bivalves, affecting filtration activity, feeding behaviour and reproductive problems
155 (Zhang et al., 2019a). Fortunately, it was demonstrated that depuration of mussels can be an
156 effective method to reduce plastic fibres (Birnstiel et al., 2019). Additionally, other
157 environmental contaminants (e.g. polychlorinated biphenyls and polycyclic aromatic
158 hydrocarbons) are affecting the health marine organisms with risk of adverse effects on
159 human health, because they can increase serious viral contaminations (Fiorito et al., 2019).

160 *3.1 Proteomic analysis for biomonitoring marine pollution*

161 Currently, the development of OMICs technologies and bioinformatics for data
162 interpretation allow advances in many scientific fields. Research marine proteomic studies
163 have been carried out with numerous contaminants or natural toxins using both field and
164 laboratory experiments (Suárez-Ulloa et al., 2013). Moreover, a proteomic approach may lead
165 to a deeper understanding of gene expression and protein translation but also protein
166 interactions or modifications (phosphorylation, denaturation, methylation, etc) in living
167 organisms (Ambrosio et al., 2018).

168 Traditional biochemical methods as well as gel-based proteomics are intensively used to
169 search protein biomarkers of marine contamination (see Table 1). In fact, in Figure 2 we can
170 see that of the articles reviewed for this study the most used proteomic methodology is the
171 2-DE method followed by MALDI TOF. However, these methods analyse a small subset of
172 proteins resulting highly biased methods with a poor quantification power (Aebersold and
173 Mann, 2016). Overall, these studies highlight the need of robust MS-based proteomics
174 methods for identification and quantification of proteins in biological tissues, therefore, their
175 application to biomonitoring the marine contamination has a significant potential. Another
176 more sensitive gel-based method such as protein labelling with fluorescent dyes (fluorescence
177 difference gel electrophoresis, DIGE) must also be considered.

178 These various proteomic methods have been applied in various toxicological studies
179 of marine organisms with animal studies being more common than plant studies due to
180 technical reasons related to the extraction of complex proteins (Figure 1). Indeed, vegetal
181 tissues have low concentrations of proteins in a high presence of interfering substances
182 (Righetti and Boschetti, 2016). Another important limitation is that only the model plant
183 species (i.e. Arabidopsis, rice, maize and wheat) have an extensive and curated protein
184 database in contrast to non-model ones (Patole and Bindschedler, 2019). Fortunately,
185 proteomic technologies and bioinformatics advances will be addressing these technical
186 problems in a short period of time.

187 Proteomic analyses have been applied to diverse marine animals such as annelids (*e.g.*
188 *Roohi-Shalmaee et al., 2019*), crustaceans (*e.g.* *Jebali et al., 2014; Vellinger et al., 2016*) and
189 fishes (*e.g.* *Borcier et al., 2019*), but the most extensive proteome have been reported in
190 molluscs for biomonitoring the aquatic environment and ecosystem health. According to
191 (Campos et al., 2012), a significant number of proteomic researches using 2-DE and MS/MS
192 showed that bivalves could be considered as aquatic environment sentinels. Therefore, it can
193 assume that the protein expression of bivalves was strongly affected by pollutants, based on
194 proteomic studies (Table 1). Thus, from the proteomic data of bivalve molluscs, it can be
195 observed that the bivalve cellular structures of different tissues are degraded by the action of
196 contaminants mainly by metal pollution. For example, Liu et al. (2012) have indicated that
197 Cd (20 and 200 $\mu\text{g L}^{-1}$ for 15 days) altered the expression of structural proteins (i.e. actin
198 and tubulin) in marine bivalves using 2-DE followed by MS/MS. In oysters, other metals
199 such as Cu, Pb and Zn until a concentration of 100 $\mu\text{g L}^{-1}$ for 4 days affected the tropomyosin
200 decreasing for 4.1, 100.9 and 6.7 times, respectively (Muralidharan et al., 2012). Moreover,
201 these metal pollutants also affected endogenous proteases as cathepsin which can degrade
202 other proteins and lead to a tissue destruction (Xu et al., 2016). For this reason, urban sewage
203 which likely includes metal pollutants also altered proteins related to cytoskeleton of oysters

204 as noted above (Flores-Nunes et al., 2015). Other non-metal pollutants as anthracene or
205 benzo α -pyrene caused alterations in structural proteins in bivalves as clam *Ruditapes decussatus*
206 (Sellami et al., 2015). All these structural proteins were detected by gel-based proteomic
207 technologies and most identified by MALDI-TOF/TOF, as discussed above.

208 Following on protein alterations in the proteome of bivalves, environmental
209 pollutants also provoke alterations in cell redox balance increasing the oxidative status of
210 proteins. The generation of free radicals such as reactive oxygen species (ROS) could cause
211 important toxicity at cellular level (Table 1). To counter this adverse effect, antioxidant
212 molecules as glutathione and other enzymes as catalase, superoxide dismutase or peroxidases
213 are expressed (Braconi et al., 2011). In *M. galloprovincialis*, a recent study has shown a level
214 change ratio for GST ([Glutathione S-transferase](#)) with a fold change of 3.5 in response to 30
215 $\mu\text{g L}^{-1}$ of polychlorinated biphenyls for 3 weeks (Ambrosio et al., 2018). Other studies have
216 revealed that these family of antioxidant enzymes can be considered as suitable proteomic
217 biomarkers (Leung et al., 2011) for the control of Cd and H_2O_2 . Other emergent
218 contaminants such as CuO nanoparticles and Cu^{+2} ($10 \mu\text{g L}^{-1}$ for 15 days) or microplastics
219 ($800 \mu\text{g L}^{-1}$ for 52 days) varied the level of proteins involved in oxidative stress, affecting its
220 cellular detoxification efficiency (Gomes et al., 2014; Green et al., 2019). Moreover, a recent
221 study with mixtures of contaminants clearly revealed the alteration of the GSTs level,
222 suggesting the activation of a defence mechanisms used to [survive in polluted environments](#)
223 (Azevedo et al., 2015).

224 Finally, another significant group of proteins altered by pollution were the proteins
225 involved in the metabolism (Table 1). Hence, proteins involved in carbohydrate, fat and
226 protein metabolism are also valuable biomarkers in bivalve organisms. Variations in
227 metabolism as a result of mechanisms of biotransformation or detoxification of pollutants
228 can be also used to monitor the contaminant trace and the ATP synthase beta subunit was
229 proposed as protein biomarker (Muralidharan et al., 2012). Heavy metal pollution caused an

230 important damage to amino acid and carbohydrate metabolism representing a 2% and 11%
231 of the total altered proteins in oysters. Particularly, the glycolytic pathway was disturbed by
232 metal pollution (Cd, Cu, Pb and Zn with a concentration of 100, 50 and 5 $\mu\text{g L}^{-1}$ for each
233 metal and during 4 days) as a result triose phosphate isomerase has been highlighted in
234 proteomic studies (Thompson et al., 2012). It has been reported that in the case of exposure
235 to CuO nanoparticles and Cu^{+2} for 15 days, most of the metabolic proteins were up-regulated
236 (i.e. ATP synthase F0 subunit 6 and cytochrome C oxidase subunit III) in the digestive gland
237 of *M. galloprovincialis* (Gomes et al., 2014). Furthermore, post-translational modifications of
238 proteins should be investigated in order to assess the functionality of proteins. At present,
239 little is known about the effect of aquatic pollution on post-translational modifications.

240 As we mentioned previously, other marine invertebrates also were studied using
241 proteomic techniques. As example, the effect of aliphatic and aromatic hydrocarbons on crab
242 *Carcinus maenas* was detected through proteins involved in several biological process as the
243 chitin catabolism, proteolysis, exoskeleton biosynthesis, protein folding, stress response and
244 transport (Jebali et al., 2014). This finding demonstrates a strong response of these chemicals
245 on the crab proteome revealing a group of 19 protein biomarkers (Jebali et al., 2014).

246 4. Biomonitoring of freshwater pollution

247 In general, the choice of aquatic bioindicator has to meet some criteria such as easy to
248 recognized, high abundance and wide distribution as well as sensitive to a wide range of
249 pollutants (Le et al., 2016). As can already be seen in Figure 1, there are far fewer
250 biomonitoring studies of freshwater pollution than of marine pollution and in fact, there is
251 no consensus on sentinel organisms in the freshwater environment. The most common
252 organisms used to biomonitoring freshwater pollution are species of fish, crustaceans and
253 molluscs but there are mainly two organisms that stand out. The bryophyte *Fontinalis*
254 *antypiretica* (Debén et al., 2015), and the zebra mussel *Dreissena polymorpha* (Binelli et al., 2015).
255 The use of *F. antypiretica* would be appealing as a simple, reliable, and economic tool for

256 biomonitoring inorganic (i.e. heavy metals and metalloids) and organic contaminants (i.e.
257 PCDD, PCDF, PAHs and PCBs). Likewise, another bryophyte species *Sphagnum palustre* was
258 recently used by Capozzi et al. (2018) for biomonitoring microplastics in freshwaters
259 environments. On the other hand, there are many published studies about zebra mussels in
260 recent years, but it is an invasive species. As recommended by Binelli et al. (2015) it should
261 not be introduced in new areas but it would be practical to use its positive peculiarities in the
262 ecosystems where it is already established or use it in laboratory experiments. But none of
263 these organisms are considered to be sentinels, as is the case with the mussel in the marine
264 studies.

265 *4.1. Proteomic analysis for biomonitoring freshwater contamination*

266 As in marine pollution, a wide range of proteomic methodologies were used up until now.
267 Proteomic technologies, including innovative combinations of different techniques, were
268 applied to pollution studies (Figure 2, Table 2). One of the first techniques was the 2-DE
269 which allows the separation of a high number of proteins followed by their identification by
270 MS spectrometry. The main drawbacks of the 2-DE technique are its time-consuming and
271 its poor reproducibility. Therefore, there is a clear trend towards gel-free alternatives such as
272 SWATH-MS, LC-MS/MS or iTRAQ as reflected in Table 2. Even though accurate and
273 sensitive mass-spectrometry-based methods enable the proteome analysis of aquatic
274 biomonitor selected, a large-scale study of proteomes requires a high-quality spectral library
275 even in the case of data-independent acquisition (DIA) strategies. Currently, data-dependent
276 acquisition (DDA)-based proteomics are the preferred methods due to the researcher do not
277 need to know about the specific proteins and the molecular mechanisms involved in the
278 biological process (Aebersold and Mann, 2016). In this sense, great efforts are also being
279 carried out to generate high-quality libraries for DIA MS in non-model organisms based on
280 fragmentations and retention time of proteins (Searle et al., 2020). When fully development,

281 the proteomic technologies will reach a maximum applicability in the biomonitoring
282 freshwater contamination.

283 At this moment, proteomic tools can provide an appealing approach to study the
284 freshwater environmental stressors on organisms. As we previously said, the species used for
285 biomonitoring freshwater pollution are highly heterogeneous and therefore we did not find
286 many articles on the same organism. Initially, plants might seem to be an ideal organism for
287 pollution biomonitoring in the rivers and lakes as we already observed with bryophytes but
288 from a proteomic perspective the situation changes. The heavy metal tolerance was deeply
289 investigated from a proteomic point out of view only in crop species, proving that metals are
290 mainly absorbed by roots and accumulate in other parts, producing important oxidative stress
291 inside the cell. In green macroalgae, it is known that the main mechanisms to counteract the
292 metal effect are the cellular inclusion of metals, synthesis of metal chelators and activation
293 of antioxidants (Moenne et al., 2016). Regarding proteomic studies, there are only a few
294 works with algae species. Recently, research conducted by Khatiwada et al. (2020) using
295 SWATH-MS, revealed that facilitator superfamily transporters, Cd/Zn transporting ATPase
296 and heavy metal transporting P1B-ATPase were overexpressed in the single-celled algae
297 *Euglena gracilis* in response to contamination of Hg, Pb and Cd in a concentration of 5-90,
298 500-10000, and 50-1000 mg L⁻¹, respectively, during 7 days.

299 As mentioned before, the most frequent aquatic animals used to monitor the
300 environmental quality are species of fish, crustaceans, and molluscs (Table 2), highlighting
301 fish in terms of the cutting edge of the proteomics tools used. From these organisms, several
302 tissues such as muscles, liver, gills, kidney, brain, or skin can be investigated, and oxidative
303 stress is one of the main mechanisms studied (Zhang et al., 2019b). Certainly, pollutants in
304 the river often induce oxidative stress, inducing devastating effect on the fish proteins. As
305 oxidative stress is an imbalance between the production of reactive oxygen species by radicals
306 and antioxidants in the cell system, cellular antioxidants such as superoxide dismutase,

307 catalase and glutathione peroxidase are often differentially expressed to achieve the balance.
308 Additionally, heat shock proteins are also involved in the regulation of the redox potential
309 via repairing damaged proteins or degrading them when the situation is irreversible. In this
310 sense, the freshwater catfish *Rita rita* has been proposed to monitor riverine pollution, due
311 to the expression of a reference proteome which included a set of heat shock proteins
312 (Mohanty et al., 2015). Moreover, the proteome of goldfish (*Carassius auratus*) subjected to
313 herbicides and fungicides (8.4 and 42 $\mu\text{g L}^{-1}$, respectively) mixture and high temperatures (22
314 and 32 °C) produced significant changes in response to cellular stress (Gandar et al., 2017).
315 In the same line, alteration of heat shock proteins and catalase was confirmed in *Gobiocypris*
316 *rarus* exposed to benzotriazole (0.05, 0.5 and 5 $\mu\text{g L}^{-1}$) (Liang et al., 2017).

317 The cellular metabolic pathways are reorganized in fish species in response to
318 inorganic and organic compounds of contaminated environments (Table 2). Within energy
319 metabolism, the main pathways studied are lipid and carbohydrate metabolism and it has
320 been reported that organic contaminants in aquatic environments could be bioaccumulated
321 in the lipids of organisms with adverse effects on transport, biosynthesis and β -oxidation of
322 fatty acids. The proteins responsible for fatty acid metabolism comprise those involved in
323 the hydrolysis of triacylglycerol, intracellular transport of fatty acids, fatty acid oxidation from
324 mitochondrial and peroxisome (Olivares-Rubio and Vega-López, 2016). These facts have
325 been proved in the proteome of white sucker (*Catostomus commersonii*) under the effects of
326 mining dust in Athabasca river (Canada) (Simmons and Sherry, 2016). Concerning
327 carbohydrate metabolism, proteins such as alpha-enolase, triose phosphate isomerase B,
328 glyceraldehyde-3-phosphate dehydrogenase and pyruvate carboxylase were altered by DDT
329 and Cd (500 and 1 mg L^{-1} , respectively) in blood cells of European eel *Anguilla anguilla*.
330 Indeed, these proteins have been point out as potential biomarkers proteomic index to detect
331 pollutants (Roland et al., 2016). In the case of rainbow trout (*Oncorhynchus mykiss*) exposed to
332 an herbicide (diquat dibromide; from 0.12 to 10 mg L^{-1} during two 24 h pulses), an

333 enrichment analysis of liver proteins differentially expressed showed that they were strongly
334 linked to glycogen degradation. A malfunction of the glycogen metabolic pathway means
335 that the prolonged swimming of the fishes without glycogen and triglycerides could lead to
336 greater predation (McCuaig et al., 2020). On the other hand, the systemic damage as the
337 result of pollutants in living organisms could be assessed studying the sub-proteome and
338 consequently, the proteomic analysis becomes focused on a particular organ tissue (Gouveia
339 et al., 2019). It has been reported that the fish neuroendocrine system is clearly affected by
340 pollutant chemicals from the environment, as well as their effect could be monitored by
341 protein biomarkers related to its system (Martyniuk et al., 2012).

342 The crustaceans are other studied group in the field of freshwater biomonitoring as
343 sentinel organism due to their sensibility for assessing the water pollution. Crustaceans offer
344 numerous advantages in terms of their small size, abundance in the freshwater ecosystem
345 and their position in the food web. Recently, it has been demonstrated that several enzymes
346 of crustaceans involved in antioxidant system response, detoxification mechanisms and
347 damage occurrence responded to polluted environments (Bertrand et al., 2018). Another
348 study analysed the impact of chlordecone, an organochlorine insecticide, on the proteome
349 of the crustacean *Macrobrachium rosenbergii* resulting a possible endocrine disruptor compound
350 in chemical concentrations of 0.2, 2 and 20 $\mu\text{g L}^{-1}$ for 30 days (Lafontaine et al., 2017). Using
351 LC-MS/MS, *Daphnia magna* was studied to assess organic contamination in freshwater. A set
352 of 288 proteins from energy metabolism and oxidative stress were altered by the impact of
353 the contamination in Gulpo river (South Korea) (Chatterjee et al., 2019). In other study
354 conducted with *Carcinus maenas*, the CBR-NHR-218 nuclear hormone receptor, two
355 components of the ABC transporters and the aldehyde dehydrogenase, were proposed as
356 biomarkers (Montes Nieto et al., 2010). As described in fish species, a polluted environment
357 can produce oxidative stress by redox unbalance. This stress caused by a polluted
358 environment in aquatic ecosystems of Doñana National Park (Spain) could be analysed by

359 means of redox proteomics using crayfish (*Procambarus clarkii*) as reported Fernández-Cisnal
360 et al. (2017), who detected cysteine oxidized forms in ferritin and nucleoside diphosphate
361 kinase. At a more advanced level, targeted quantitative proteomics by mass spectrometry can
362 also provide valuable information for detecting a mixture of pollutants in a freshwater
363 ecosystem. Based on current protein databases, a selected reaction monitoring mass
364 spectrometry-based methodology in amphipod crustaceans corroborated the methodology
365 to assess the aquatic contamination (Gouveia et al., 2017).

366 Finally, for biomonitoring of environmental quality, bacteria are not very common in
367 proteomic studies and they are focused on the use of microorganisms to destroy or reduce
368 the environmental pollutants through bioremediation. In this sense, recent proteomic studies
369 could provide a better understanding of molecular mechanisms to improve efficiently the
370 bio-removal of pollutants (Wei et al., 2019; Yu et al., 2020).

371 **5. Final remarks and future trends**

372 There is a rising concern about water pollution caused by industrial and agricultural activities.
373 These scenarios are in constant evolution and the type of the contaminants is constantly
374 changing. Emerging contaminants and new threats require new approaches, identifying novel
375 biomarkers in aquatic organism. As shown in the Figure 3, the application of proteomics is
376 increasingly relevant in aquatic pollution searching protein biomarkers in novel sentinel
377 organisms. Additionally, protein biomarkers represent a valuable tool for early detection of
378 pollutant exposure and even early effect evaluation. The proteomic approach is one of the
379 most dynamic and fast developing field to analyse and decipher protein expression. **However,**
380 **the establishment of biomonitoring protocols implies a great effort due to the seasonal and**
381 **spatial variability of communities as well as individual variability. Past researches have taken**
382 **the initial steps using lab experiments to create models, but more complex systems with**
383 **interactions among pollutants and uncontrolled environmental conditions must be**
384 **considered in the future.**

385 Nevertheless, one of the main difficulties is the lack of a curated protein database for
386 these living organisms. To overcome this limitation, new proteomic mass spectrometry
387 technologies are being developed to detect and quantify the proteins increasing the precision
388 and accuracy. In recent years, DNA sequencing towards precise genomes are implementing
389 proteome data building an accurate reference protein database. It is becoming increasingly
390 the number of non-model aquatic organisms with enough proteomic data, thereby opening
391 new avenues for proteomic approach.

392 In the following years most of studies should be focused on the study of protein
393 isoforms, post-translational modifications and protein interactions which could reveal
394 further information on the stress response to chemical pollution. The analysis of alternative
395 protein isoforms and protein interactions would provide insight into the relation between
396 proteins and their functions. Additionally, post-translational modifications of proteins,
397 particularly phosphorylation, which play key roles in the regulation of protein functions and
398 signalling networks of biological systems is therefore a major challenge.

399

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- 659
- 660

661 **CAPTIONS TO FIGURES**

662 **Figure 1.** Number of articles published during the last 10 years using plants (A) and animals
663 (B) for marine  and freshwater  pollution biomonitoring; those studies that employed
664 proteomic tools are highlighted .

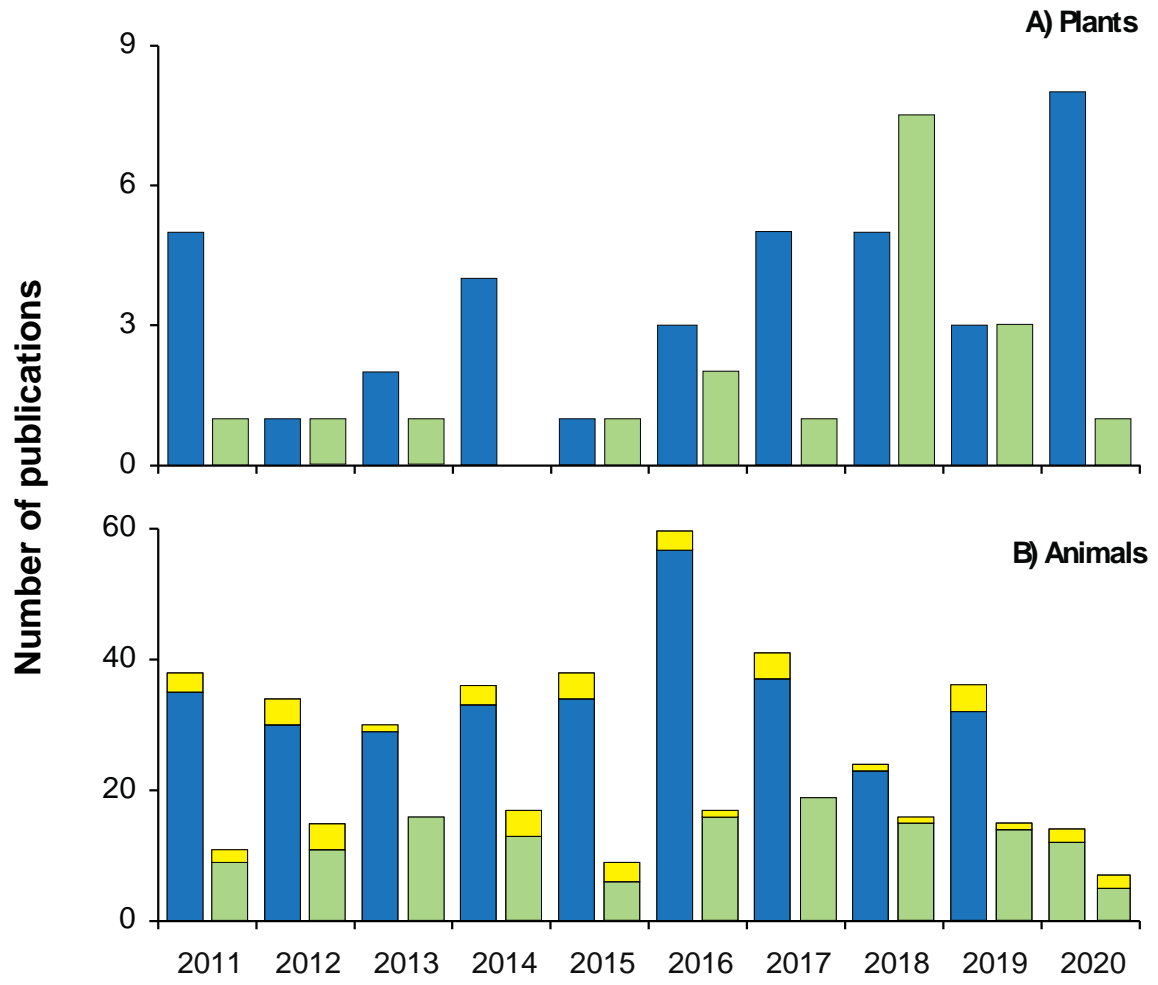
665 **Figure 2.** Bar chart highlighting proteomic methodology used in revised studies about
666 pollution biomonitoring for aquatic environment from 2010 to 2020.

667 Figure acronyms: LC-Q-TOF=Liquid chromatography-quadrupole-time of flight; SWATH-
668 MS= Sequential window acquisition of all theoretical fragment ion spectra mass
669 spectrometry; SDS-PAGE= Sodium dodecyl sulfate-polyacrylamide gel electrophoresis; RP-
670 HPLC= Reverse Phase High Performance Liquid Chromatography; LFQ= Relative label-
671 free quantification of shotgun; 2D-UPLC= Two dimensional by ultra performance liquid
672 chromatography; PLGS= Protein Lynx Global Server; 2D-DIGE= Two-dimensional
673 difference gel electrophoresis; 2-DE= Two-dimensional gel electrophoresis; LC-MS= Liquid
674 chromatography mass spectrometry; Nano-LC-MS/MS= Nano- liquid chromatography-
675 tandem mass spectrometry; iTRAQ= Isobaric tags for relative and absolute quantitation;
676 MALDI TOF/TOF= Matrix-assisted laser desorption ionisation - tandem mass
677 spectrometry (TOF)

678

679 **Figure 3.** Advantages and drawbacks of pollution biomonitoring from a proteomic point of

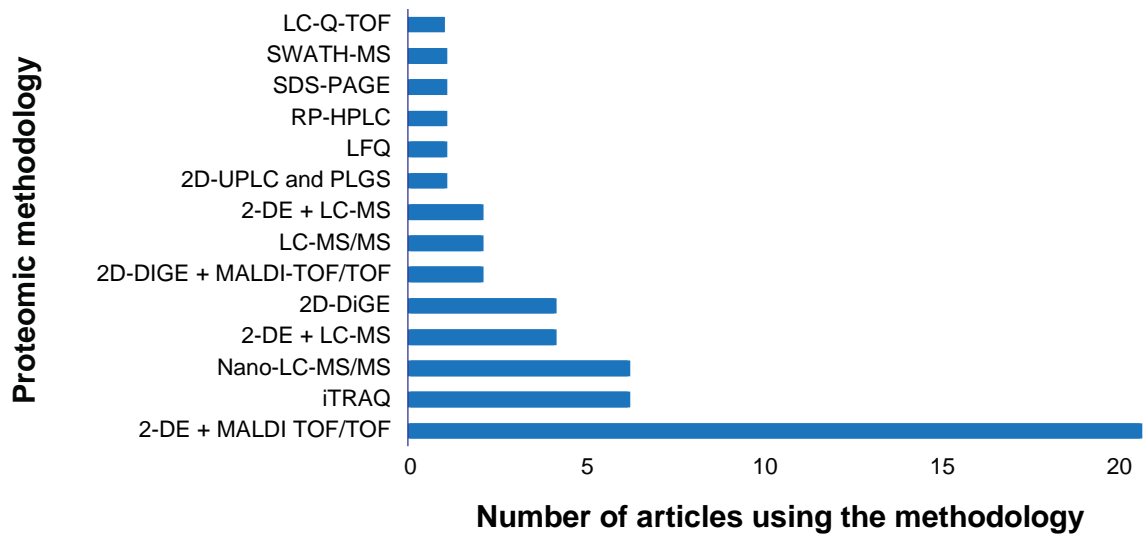
680 view



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2
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Figure 1

1



2

3 **Figure 2**

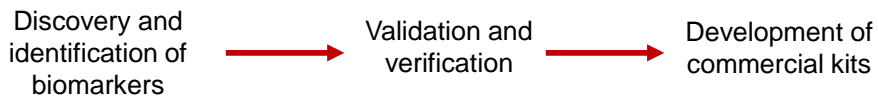
4

Advantages and drawbacks of a pollution biomonitoring using a proteomic approach

- ✓ Identification of **new protein biomarkers**
 - Emerging contaminants
 - New biomonitors
 - New scenarios
- ✓ **Early detection** of pollutant exposure / biological effect
- ✓ In-depth **knowledge of molecular mechanisms** altered by pollution

- ✗ **Additional effort** would be needed
 - Optimization of proteomic techniques** for new tissues of biomonitor species
 - Curated protein databases** for non-model organisms is not available yet

- ✗ **Intensive research** to reach a commercial kit of biomarkers, following the detailed procedure:



1

2 **Figure 3.**

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