

1 **Evaluation of the efficacy of two postweaning colibacillosis vaccines in a field herd with PRRS**
2 **circulation during postweaning stage.**

3 Gonzalo López-Lorenzo¹, Alberto Prieto¹, José Manuel Díaz-Cao^{1*}, Cynthia López-Novo¹, David
4 García-Dios¹, Ceferino López¹, Rosario Panadero¹, Antonio Iglesias², Pablo Díez-Baños¹, Gonzalo
5 Fernández¹

6 ¹ Department of Animal Pathology (INVESAGA Group), Faculty of Veterinary Sciences, Campus
7 Terra, Universidade de Santiago de Compostela, 27002 Lugo, Spain

8 ² Department of Anatomy, Animal Production and Veterinary Clinical Sciences. IBADER. ,
9 Universidade de Santiago de Compostela, 27002 Lugo, Spain

10

11 *Corresponding author: José Manuel Díaz-Cao

12 Tel.:+34 982 822127 E-mail: josemanueldiaz.cao@usc.es

13 **ABSTRACT**

14 Postweaning diarrhea (PWD) and PRRS are two major concerns in swine production, which
15 association has not been consistently explored. In the current scenario of restrictions in the use of
16 antibiotics and ZnO, vaccination is more relevant to control PWD, but PRRS virus circulation may
17 compromise the immune protection conferred by postweaning colibacillosis vaccines. We evaluated
18 the efficacy of two postweaning colibacillosis vaccines (parenteral and oral) in a commercial herd
19 affected by an outbreak of PWD and with PRRS circulation in postweaning. Five groups were
20 studied during the postweaning period: one control (Group 1) and four vaccinated: two with each
21 postweaning colibacillosis vaccine administered alone (Groups 2 and 3) or with sow vaccination
22 against PRRS (Groups 4 and 5). We evaluated the effects on piglet weight, average daily weight
23 gain and in the percentage of piglets with diarrhea, its duration, lethality and mortality. PRRS
24 viremia and anti-PRRS antibodies were evaluated by qPCR and ELISA. Regarding control group,
25 colibacillosis vaccination generally improved most of the measured parameters; but significant
26 improvements were only observed in Groups 4 and 5 ($p < 0.05$). Moreover, cases of diarrhea
27 occurred at different ages: in Groups 2 and 3 the peak of cases occurred just after ZnO was removed
28 from the feed compared to Group 1, while in Groups 4 and 5 no peak was observed. This suggests
29 that postweaning colibacillosis vaccination may be compromised by the PRRS circulation. In PRRS
30 endemic herds an effective protection against PWD through vaccination may require PRRS
31 vaccination to obtain a better performance.

32 **KEYWORDS:** diarrhea, *E. coli*, postweaning colibacillosis, PRRS, swine, vaccination, ZnO.

33 INTRODUCTION

34 Swine postweaning diarrhea (PWD) is a multifactorial disease responsible of significant economic
35 losses worldwide as it is related with increased mortality, animal weight loss and growth retardation
36 (Fairbrother et al., 2005). In addition, the occurrence of PWD also presents a risk for the emergence
37 of resistances as farms with this problem present an increased use of antibiotics due to treatments.
38 (García-Meniño et al., 2021). PWD is usually the result of environmental and husbandry factors that
39 facilitate the infection with specific pathogens around weaning. Many pathogens can be involved in
40 PWD, but Enterotoxigenic *Escherichia coli* (ETEC) is the most important among all of them
41 (Fairbrother et al., 2005). In order to cause diarrhea, ETEC must present two specific virulence
42 factors: fimbriae adhesins, which facilitate the bacteria attachment to the enterocytes (e.g., F4, F18
43 and less frequently F5, F6 or F41); and enterotoxins, which are responsible of the toxic effect and
44 disrupt the intestinal homeostasis (heat-labile toxin (LT), heat-stable toxin a (STa) and heat-stable
45 toxin b (STb)) (Fairbrother et al., 2005; Fairbrother and Nadeau, 2019). When ETEC is causing
46 PWD, it generally affects to a high proportion of piglets, which typically show grey or yellowish
47 watery diarrhea along with a characteristic smell around the 2-3 weeks after weaning. The disease
48 usually lasts about one week, although sudden deaths can also be observed (Luppi, 2017).
49 Secondary and immunosuppressive infections may also exacerbate the course of this disease (Drew,
50 2000; Segalés et al., 2004).

51 Several control strategies against PWD have been proposed: increasing the weaning age and
52 performing creep-feeding during the lactation period, increasing the fiber levels and reducing the
53 protein in the piglet diet, water acidification, the addition of organic acids or prebiotics to the feed,
54 etc. (Zentek et al., 2012; McLamb et al., 2013; Jha and Berrocso, 2016; Dubreuil, 2017; Rhouma
55 et al., 2017; Lee et al., 2021). However, the preventive feed medication of piglets with zinc oxide
56 (ZnO) has been the most successful and extended measures to control the proliferation of ETEC and
57 incidence of PWD. Levels up to 3,000 parts per million (ppm) of ZnO in feed have shown to reduce
58 clinical development of diarrhea, the mortality rate and improve growth (Luppi, 2017). However,

59 since July 2022 the use of ZnO as a feed additive in the European Union is prohibited due to
60 environmental concerns (The European Medicines Agency, 2017). These new restrictions may
61 increase the incidence and mortality rate of PWD in swine herds, as well as make other secondary
62 intestinal disorders more important.

63 Despite different postweaning colibacillosis vaccines are commercially available, vaccination of
64 piglets has been limitedly used on field due to the success provided using ZnO (Melkebeek et al.,
65 2013), but this practice may gain now more importance under the new restricted scenario. However,
66 the efficacy of vaccination against postweaning colibacillosis has been limitedly assessed and
67 mostly in controlled conditions (Van der Stede et al., 2003; Fairbrother et al., 2017; Nadeau et al.,
68 2017; Vangroenweghe and Thas, 2020; Correa et al., 2022; Ramis et al., 2022; Vangroenweghe and
69 Boone, 2022), so studies in field conditions are highly required, for example, to evaluate
70 interferences with other endemic infections. Most of the current commercial postweaning
71 colibacillosis vaccines are for parenteral or oral administration and include the most important
72 adhesins (F4 and F18). Parenteral vaccines stimulate the active immunity in piglets at systemic
73 level, while the latter ones mainly active the production of secretory IgA antibodies at intestinal
74 level (Melkebeek et al., 2013). The effectiveness of these vaccines against colibacillosis in field
75 conditions can be compromised by low immunity status in the piglets or by the coinfection with
76 immunosuppressive infections. In this regard, Porcine Reproductive and Respiratory Syndrome
77 (PRRS) virus is an immunosuppressive infection that circulates endemically in most of the pork-
78 producing countries and constitutes one of the major economic limiting factors in swine industry.
79 The infection with PRRS virus may interfere in the development of an adequate immunity against
80 other infections, reducing the effectiveness of the vaccines (Sang et al., 2011; Lunney et al., 2016).
81 Moreover, it has been reported that PRRS infection induces intestinal damage, shortening the villus
82 height and altering the jejunal function (Escobar et al., 2006; Helm et al., 2020); and that it causes
83 the disruption of intestinal barrier by weakening tight junction barrier integrity (Zhao et al., 2021),
84 which could also aggravate enteric process such as postweaning colibacillosis.

85 Therefore, vaccination against ETEC *E. coli* is expected to gain demand after the prohibition of
86 ZnO, but questions regarding the on-field efficiency of the current vaccines are still unresolved,
87 especially when circulation of PRRS is happening simultaneously. Hence, we carried out an
88 experimental study in the postweaning stage of a herd affected by an outbreak of PWD and with
89 circulation of PRRS to evaluate the effect on the production parameters and mortality of two
90 commercial vaccines against postweaning colibacillosis, applied either alone or together with a
91 PRRS vaccination program.

92 **MATERIAL AND METHODS**

93 **Farm description and nutrition**

94 The present study was performed during 2020 on a conventional farrow-to-finish pig farm with 150
95 sows managed in a one-week batch-management system. Piglets were weaned at 21 days of age and
96 housed in post-weaning facilities during approximately six weeks. Pigs were placed in pens with
97 total-slatted floor in groups of 13-14 piglets. Water and food were available *ad libitum* following a
98 3-phase feeding strategy: from 21 to 28 days old piglets were fed with a lacto-initiator (the same
99 that had been used for creep-feeding), from 29 to 42 days a weaning starter supplemented with ZnO
100 (3.100 ppm), and from 43 days onwards using a starter. Ventilation was performed by an automated
101 air fan located in the roof of the room and fresh air entered the room directly from outside.

102 **Herd health situation**

103 The herd presented a history of PRRS circulation in postweaning piglets. One year before starting
104 the present study, the herd had stopped its vaccination program against PRRS virus at weaning, so
105 piglet vaccination only include vaccination against *Mycoplasma hyopneumoniae* and Porcine
106 Circovirus Type 2 at 15 and 28 days old, respectively; piglets were never vaccinated against *E. coli*.
107 Regarding the breeding-herd population, it was free of PRRS and had not been vaccinated before
108 against this virus; the sow vaccination schedule includes a vaccination against *E. coli* and
109 *Clostridium perfringens* Type C one month before farrowing.

110 Approximately 2-3 months before the beginning of the study, the farm suffered an outbreak of
111 PWD affecting to piglets between 5 and 6 weeks old. Accordingly, rectal swabs and serum samples
112 were taken from affected piglets (n=5) and analyzed by qPCR using a digestive diagnostic panel
113 (F4, F5, F6, F41, F18, STa, STb, LT, Stx2, *eae*, AIDA and EAST genes from *E. coli*, *Salmonella*
114 spp., Rotavirus A and Porcine Epidemic Diarrhea) and to detect PCV2 and PRRS viremia. Results
115 from rectal swabs were positive to *E. coli* and included the detection of F4, F18, STa, STb, LT, *eae*,
116 AIDA and EAST genes, as well as Rotavirus A (Cq 23.10 to 32.20). Regarding the results from
117 serum samples, PRRS viremia was detected in all serum samples (Cq 26.11 to 30.52), while PCV2
118 viremia was discarded.

119 **Study design**

120 We carried out different vaccination schemes against PRRS and ETEC in five different groups of
121 pigs (Table 1). The groups were created from five consecutive batches, so all the animals in each
122 batch were assigned to each group. All the protocols and procedures followed during the study were
123 approved by the bioethics committee of the University of Santiago de Compostela (project
124 identification code: 2019-CE194). Individual piglets with clinical signs of PWD (watery feces and
125 clinical signs of dehydration) were treated with a commercial injectable sulfadoxine-trimetoprim
126 and were fasting while diarrhea was evident. Other disorders were also treated according to
127 veterinarian recommendations.

128 Data collection and laboratory analysis

129 Piglets were individually ear tagged at weaning and weighted at 21, 28, 42 and 63 days old. The
130 average daily weight gain (ADWG), food intake (total kg food/piglet), and feed conversion rate
131 (FCR) (total kg food/total kg piglet weight gain) of piglets were calculated for each feeding period:
132 lacto-initiator (between 21 to 28 days old), weaning starter (between 29 to 42 days old), starter
133 (between 43 to 63 days old); and for all the study period (between 21 to 63 days old).

134 Measurements from animals that dead during the study were registered if the piglet had started the

135 corresponding feeding period up to the moment of death and using their dead weight when was
136 necessary for calculations.

137 Clinical signs of diarrhea (grey or yellowish watery feces) and the presence of dead were evaluated
138 daily from 21 to 63 days old. We registered the week when clinical signs began and the duration of
139 the diarrhea (days with clinical signs); dead piglets were individually weighed and registered in the
140 post-weaning corresponding week.

141 The presence of antibodies and viremia against PRRS virus was monitored in Groups 1 and 4, as
142 control and piglets from PRRS vaccinated sows, respectively. Serum samples were collected at 10,
143 25, 41 and 58 days old from a minimum of 20 random piglets from each group. These samples were
144 frozen at -80 °C until laboratory analysis. The serum samples were analyzed to detect the presence
145 of antibodies against PRRS virus using the commercial kit Ingezim PRRS Universal (INGENASA,
146 Madrid, Spain). A serum sample was considered positive to PRRS virus IgG antibodies if the
147 ELISA sample/positive (S/P) ratio was higher than 0.25, then PRRS virus antibody titer was
148 calculated according to manufacturer's instructions. For titer calculation of negative samples, a S/P
149 ratio of 0.20 was considered. To detect viremia, serum samples at each sampling moment were
150 pooled (5 pigs/pool) and pools were analyzed by qPCR. RNA extractions were performed using the
151 commercial kit NucleoSpin[®] RNA (Macherey-Nagel, Düren, Germany) following manufacturer's
152 instructions, using 200 µL of each blood pool as starting material and collecting the extracted RNA
153 in 60 µL of molecular water. The isolated RNA was analyzed by qPCR using the commercial kit
154 VetMAX PRRSV EU/NA (ThermoFisher Scientific, Waltham, Ma, USA). A sample was
155 considered positive when $Cq < 40$. qPCR reactions were run on an Applied Biosystems ABI Prism
156 7500 thermocycler (ThermoFisher Scientific, Waltham, MA, USA).

157 **Statistical Analysis**

158 A linear regression analysis was used to evaluate the differences in piglet weight between groups at
159 each sampling point. Only the piglets with all the weight measures were included for this analysis

160 excepting the weight evaluation at the beginning of the study, in which all piglets were included.
161 Linear regression analysis was also performed to evaluate differences between groups in the
162 ADWG in each feeding period as well as in all the study, the number of days with clinical diarrhea
163 and the PRRS antibody titer at each sampling age.
164 We used a logistic regression to evaluate the presence of differences in the proportion of piglets
165 with diarrhea and the accumulated mortality by group of vaccination.
166 Data analysis was performed in R v. 4.1.1 (R Core Team, 2021) (Vienna, Austria). In all analyses a
167 $p < 0.05$ was considered as significant.

168 **RESULTS**

169 **Animal weight and ADWG**

170 The weight of the piglets and the ADWG could be assessed in 375 out of 414 animals. The
171 remaining ones were excluded due to death. Regarding weight, no significant differences were
172 found in piglets at 21 and 28 days old between the groups of vaccination, but they were detected in
173 older piglets (Figure 1). In this case, animals with double protection against *E. coli* and PRRS
174 (Group 5) were the only ones that showed significantly higher weights than the control group
175 (Group 1), at 42 or at 63 days old. In contrast, animals with only *E. coli* IM vaccination (Group 2)
176 showed a worse final weight than the control group.

177 Regarding ADWG, we found statistically significant differences among groups in all the feeding
178 periods (Figure 2). It must be highlighted that during the first postweaning week the ADWG in
179 groups with PRRS vaccination (Group 4 and 5) was significantly higher than the control and the
180 other groups. Thereafter, the ADWG was not significantly higher in any vaccination group than in
181 the control group.

182 Results for food intake and FCR for each group are showed in Table 2. The most remarkable
183 observation occurred during the period of feeding with lacto-initiator, in which upon of having a

184 similar food intake, the FCR was slightly better in *E. coli* vaccinated groups and notoriously better
185 in groups with PRRS vaccination.

186 **Postweaning diarrhea, mortality and lethality**

187 Compared to the control group, the vaccination programs significantly decreased the cumulative
188 proportion of piglets affected by PWD in Groups 3 to 5, especially in Groups 4 and 5: Group 3 (OR
189 = 0.394; 95% CI = 0.161 – 0.098; $p = 0.031$), Group 4 (OR = 0.101; 95% CI = 0.016 – 0.365; $p =$
190 0.003) and Group 5 (OR = 0.083; 95% CI = 0.013 – 0.299; $p = 0.001$) (Table 3). In contrast, Group
191 2 did not show a significant reduction of the proportion of piglets affected by PWD (OR = 0.903;
192 95% CI = 0.426 – 1.891; $p = 0.788$). The clinical duration of PWD was also reduced in all the
193 vaccinated groups, although only significantly in Groups 2 and 3 compared to Group 1 (Table 3).
194 However, Groups 4 and 5 only presented two diarrhea clinical cases.

195 In addition, while most of the affected piglets from Group 1 appeared in the three first postweaning
196 weeks, the pattern was different in Groups 2 and 3, and the peak of cases was delayed to the fourth
197 postweaning week, coinciding with the change of weaning starter feed with ZnO to the starter feed.
198 Groups 4 and 5 did not suffer that peak after this feeding change (Figure 3).

199 Mortality rates were generally lower in vaccinated groups, though the difference was only
200 significant for Group 4 (OR = 0.261; 95% CI = 0.058 – 0.852; $p = 0.042$). No significant
201 differences were observed between vaccinated groups (Table 3). However, the number of casualties
202 attributed to PWD in vaccinated piglets was low (ranging 1-2), as well as the lethality. The
203 exceptions to these observations occurred in Groups in which denominators were very small (< 3),
204 so the percentages may be unreliable. Due to the small number of observed cases, no statistical
205 analysis was performed for the % of mortality due to diarrhea and to % of lethality.

206 **PRRS monitoring**

207 We did not detect PRRS viremia until 25 days of life (three positive pools, Cq ranged from 31.86 to
208 34.53) in piglets from unvaccinated sows. In piglets from vaccinated sows, the first detection

209 occurred later, at 41 days old (three positive pools, Cq ranged from 29.80 to 34.41). After that,
210 PRRS virus circulation was detected in both groups with similar Cq values until the end of the study
211 (Figure 4).

212 PRRS serology differed between those piglets from vaccinated and from unvaccinated sows (Figure
213 4). At the beginning of the monitoring, the piglets from vaccinated sows showed a significantly
214 higher level of anti-PRRS antibodies ($p = 0.016$) than those from unvaccinated sows, however, at
215 25 and 41 days of life the levels were similar ($p > 0.05$). Instead, at 58 days old the level of anti-
216 PRRS antibodies was significantly higher in those piglets from unvaccinated sows ($p = 0.036$) than
217 those from vaccinated ones.

218 **DISCUSSION**

219 In the present study we evaluated the efficacy of two postweaning colibacillosis vaccines applied
220 alone or combined with a reinforcement of PRRS immunity in a farm affected by an outbreak of
221 PWD and PRRS circulation during postweaning. Upon the technification in swine production
222 during the last decades, both PWD and PRRS remain as two of the major concerns in this livestock
223 sector. However, a close association between PWD and PRRS was not historically considered. In a
224 scenario of increasing restrictions on the use of antibiotics and products such as ZnO, which are
225 widely used for the control of PWD, preventive measures, such as immunoprophylaxis, gain special
226 relevance. However, the efficacy of vaccines may be limited when immunosuppressive agents such
227 as PRRS virus are circulating (Drew, 2000; Kitikoon et al., 2009). Thus, the combination of
228 vaccination schedules for *E. coli* and PRRS virus could be beneficial to improve the consequences
229 of PWD but, as far the authors known, this has not been explored in the scientific literature.

230 Our results confirm that postweaning colibacillosis vaccination aids to reduce the incidence and
231 magnitude of PWD, though differently depending on if it was performed in groups with PRRS sow
232 vaccination. We generally found fewer animals affected by diarrhea and for less time in groups with
233 piglets vaccinated against postweaning colibacillosis. This vaccination also reduced the impact of
234 PWD in the survival of piglets. This is evidenced first, by a reduction of the mortality rate in

235 vaccinated groups, which was statistically significant in Group 4, and second, by a general
236 reduction of the mortality attributed to diarrhea, except in Group 4, in which the result could be an
237 artifact due to the very small number of casualties (Table 3). Lethality also tended to be lower,
238 though we could not statistically assess it due to the low number of counts. Groups with PRRS-
239 vaccination reported more lethal diarrhea cases, but considering that only 1-2 animals were
240 affected, sporadic conditions such as bad vaccinations, weak animals, etc. cannot be ruled out. It
241 must also be noted that the number of antibiotic treatments is likely to be lower in vaccinated
242 groups due to the lower incidence of PWD cases. However, treating PWD with antibiotics could
243 also secondarily reduce the incidence of other bacterial diseases. Thus, the constant reduction in
244 antibiotic treatments in consecutive batches due to vaccine efficacy might be correlated with a
245 sustained intensification in the direct transmission of some secondary infections, experiencing in
246 vaccinated groups an increase in their incidence and/or mortality rates. For example, Group 5
247 presented a similar mortality to the control group, but the percentage of animals affected by PWD
248 was markedly lower (2.33% vs 22.22%). Most of casualties in this group consisted of sudden deaths
249 with polyserositis and *Streptococcus suis* was diagnosed as the responsible. The increase of *S. suis*
250 incidence may have been favored by a reduced antibiotic treatment against PWD used in this
251 vaccinated group. Nevertheless, changes in the incidence of other diseases after ETEC vaccination
252 are not well clarified. For example, Vangroenweghe and Boone, 2022 reported the exact opposite
253 situation, a lower incidence of streptococcal meningitis in pigs vaccinated for ETEC. Overall,
254 further studies are needed to assess the incidence of other pathologies after controlling PWD.

255 Postweaning colibacillosis vaccination was also positive to improve the productive parameters in
256 most of the analyzed scenarios. First, it is notable the FCR values observed in the first postweaning
257 week: in Group 1, despite the ingestion of lacto-initiator, the whole of piglets lost weight probably
258 as consequence of PWD outbreak and PRRS circulation, in addition, fasting animals due to PWD
259 probably contributed to magnify this value; in Groups 2 and 3 the postweaning colibacillosis
260 vaccination was able to reduce the magnitude of PWD although the piglets were not able to convert

261 the feed energy in meat, which explains the high values observed in both groups; in Groups 4 and 5,
262 with the reinforcement against PRRS infection, the FCR achieved normal values. Second, apart
263 from Group 2, food intake per piglet for each feeding phase and ADWG in the lacto-initiator and
264 weaning starter periods were better in all the vaccinated groups, and remarkably in Groups 4 and 5,
265 than in the control group. These results are consistent with those reported by Correa et al. (2022),
266 who indicated that vaccination against postweaning colibacillosis can promote gut health specially
267 in the two first postweaning weeks. Third, same as these authors, we observed that unvaccinated
268 piglets used more efficiently the starter feed than vaccinated piglets. We hypothesize that the lower
269 FCR may be mainly related with the absence of immune response in unvaccinated piglets. The
270 vaccination involves an energy consumption in the development of antibody and mucosal
271 immunity, which may be remarkable in oral vaccinations (Van der Weken et al., 2021); thus, it
272 could influence in the lower FCR observed in control group. In addition, the higher antibiotic
273 treatment requirement in this group could reduce the influence of secondary infections and
274 contribute to the better FCR observed in unvaccinated piglets. Regarding Group 2, it must be noted
275 that 8 – 10 % of the piglets presented a slightly swollen neck. This has been notified as a possible
276 secondary effect of the vaccine by the manufacturer; however, we did not observe it in the other
277 group that used the same vaccine. We hypothesize that repeated injections in the neck may have
278 favored this finding, as it was more frequent in animals treated by the intramuscular route due to
279 PWD. Despite being a minor lesion, it could have caused some discomfort that had an impact on the
280 food intake and therefore, on the final weight of the piglets in this group.

281 We also found a peak in the percentage of piglets with PWD just in the first week without ZnO in
282 the feed but only in groups without PRRS vaccination (Figure 3). This phenomenon has been also
283 notified by other authors, even naming it as “post-ZnO diarrhea” (Vangroenweghe and Thas, 2020).
284 On the contrary, in groups with PRRS vaccination, no piglets showed diarrhea from the third
285 postweaning week onwards. Thus, in field conditions, the vaccination against postweaning

286 colibacillosis itself may not be enough to totally prevent the diarrhea when the ZnO at therapeutic
287 levels withdraws from the feed.

288 The improvement observed with colibacillosis vaccination was significantly better when both types
289 of *E. coli* vaccines were applied to piglets from PRRS vaccinated sows. This suggests that PRRS
290 infection interferes with the vaccination against *E. coli* and compromises the success of the
291 measure. Regardless the type of *E. coli* vaccine administered, we detected viral circulation later in
292 the groups with PRRS vaccination of sows (around three weeks postweaning) along with a
293 significant reduction in the proportion of piglets with diarrhea. The later circulation of PRRS in
294 piglets is a consequence of the sow's vaccination. It is known that the vaccination and revaccination
295 of sows with the employed PRRS vaccine induces a booster effect in their immunity (Díaz et al.,
296 2013; Geldhof et al., 2013), and a higher level of PRRS virus maternal derived antibodies in their
297 offspring, as it was observed in the present study. This can result in high titers of neutralizing
298 antibodies from passive transfer that contribute to a low susceptibility of piglets to PRRS and to a
299 delayed age of infection (Lopez et al., 2007; Hsueh et al., 2021; Martín-Valls et al., 2022), which is
300 consistent with our results.

301 The present study suggests that the PRRS passive protection during the first postweaning weeks
302 prevents piglets from PRRS circulation and, as consequence, favors the development of immune
303 system which may have important effects in terms of prevention of PWD and in productive
304 parameters. Thus, this delay of PRRS circulation may explain the lower percentage of piglets with
305 diarrhea in groups with PRRS vaccination (Groups 4 and 5) regarding groups without PRRS
306 vaccination (Groups 2 and 3). As postweaning colibacillosis vaccines are usually administered
307 around weaning, delaying the circulation of PRRS virus also delays any potential interference that
308 the infection may have with the colibacillosis vaccination and the age in which the virus causes
309 immunosuppression and intestinal damage (Escobar et al., 2006; Helm et al., 2020; Zhao et al.,
310 2021). Piglets that are susceptible of PWD at an older age are less likely to suffer from severe
311 clinical signs as they have a more mature immune system, as well as a greater ability to feed, which

312 implies having more energy reserves and, therefore, coping better with PWD (Thomson and
313 Friendship, 2019; Ming et al., 2021). Moreover, it also should be highlighted that this delay in the
314 PRRS circulation also entails a reduction in the risk of iatrogenic transmission of this virus, which
315 was remarkable in those groups from PRRS vaccinated sows (Groups 4 and 5). In these groups, the
316 need of antibiotic administration to treat PWD was considerably reduced and, secondarily, can
317 contribute to reduce PRRS incidence in postweaning.

318 Overall, our results show that the performance of colibacillosis vaccination can be influenced by the
319 existence of other active infections. In particular, farms with endemic PRRS and unstable. If the
320 virus cannot be controlled, delaying PRRS infection can be key to ensure the *E. coli* vaccines can
321 confer protection against clinical PWD. Based on our results, vaccination against both pathogens
322 seems to protect piglets specially during the first postweaning weeks. Thus, a period of at least 2-3
323 weeks between completing the postweaning colibacillosis vaccination schedule and becoming
324 PRRS infected seems to be required to guarantee that these vaccines provide an adequate immune
325 protection. Further studies are required to characterize more in detail this observation as the demand
326 and use of vaccines against postweaning colibacillosis seems that will increase in the forthcoming
327 years.

328 **CONCLUSION**

329 The employed vaccines against postweaning colibacillosis reduced the lethality and the duration of
330 PWD and tended to improve the production parameters. However, its effectiveness is restricted by
331 the PRRS circulation which, at same time, seems to contribute to the appearance of clinical cases of
332 PWD. The reinforcement in the piglet immunity against PRRS through sow vaccination seems
333 required and beneficial to optimize the protection conferred by postweaning colibacillosis
334 vaccines in herds with PRRS circulation during the postweaning stage.

335 **FUNDING**

336 This research was supported by the Programme for Consolidating and Structuring Competitive
337 Research Groups (GRC2019/04; Xunta de Galicia, Spain) and by a postdoctoral grant to G. López-
338 Lorenzo (Campus de especialización Campus Terra, Universidade de Santiago de Compostela).

339 **INSTITUTIONAL REVIEW BOARD STATEMENT**

340 The procedures which involved live animals (vaccination and blood sampling) were approved by
341 the bioethics committee of the University of Santiago de Compostela (project identification code:
342 2019-CE194).

343 **DATA AVAILABILITY STATEMENT**

344 The data are available from the corresponding author upon reasonable request.

345 **ACKNOWLEDGMENTS**

346 We are very grateful for the collaboration and willingness of the farmers during the sampling
347 process and data collection, as part of study has been carried out during the Covid-19 pandemic.

348 **DECLARATION OF COMPETING INTEREST**

349 The authors declare that they have no known competing financial interests or personal relationships
350 that could have appeared to influence the work reported in this paper.

351 **DECLARATION OF GENERATIVE AI IN SCIENTIFIC WRITING**

352 The authors declare that they did not use generative artificial intelligence (AI) and AI- assisted
353 technologies in any part of the study development and in the paper redaction.

354 **REFERENCES**

- 355 Colidex-C, 2022. Agencia Española de Medicamentos y Productos Sanitarios. URL
356 https://cimavet.aemps.es/cimavet/pdfs/es/ft/3450+ESP/FT_3450+ESP.pdf (accessed 10.14.22).
- 357 Coliprotec F4/F18. Agencia Española de Medicamentos y Productos Sanitarios. URL
358 https://cimavet.aemps.es/cimavet/pdfs/es/ft/EU%25402%254016%2540202%2540001/FT_EU
359 [-2-16-202-001.pdf](https://cimavet.aemps.es/cimavet/pdfs/es/ft/EU%25402%254016%2540202%2540001/FT_EU-2-16-202-001.pdf) (accessed 10.14.22).

- 360 Correa, F., Luise, D., Amatucci, L., Palumbo, F., Viridis, S., Negrini, C., Clavenzani, P., Vecchi, M.,
361 Mazzoni, M., Bosi, P., Trevisi, P., 2022. Effect of an *Escherichia coli* F4/F18 bivalent oral live
362 vaccine on gut health and performance of healthy weaned pigs. *Animal* 16, 100654.
363 doi:10.1016/j.animal.2022.100654
- 364 Díaz, I., Gimeno, M., Callén, A., Pujols, J., López, S., Charreyre, C., Joisel, F., Mateu, E., 2013.
365 Comparison of different vaccination schedules for sustaining the immune response against
366 porcine reproductive and respiratory syndrome virus. *Vet. J.* 197, 438–444.
367 doi:10.1016/j.tvjl.2013.02.008
- 368 Drew, T.W., 2000. A review of evidence for immunosuppression due to Porcine Reproductive and
369 Respiratory Syndrome Virus. *Vet. Res.* 31, 27–39. doi:10.1051/vetres:2000106
- 370 Dubreuil, J.D., 2017. Enterotoxigenic *Escherichia coli* and probiotics in swine: What the bleep do
371 we know? *Biosci. Microbiota, Food Heal.* 36, 75–90. doi:10.12938/bmfh.16-030
- 372 Escobar, J., Toepfer-Berg, T.L., Chen, J., Van Alstine, W.G., Campbell, J.M., Johnson, R.W., 2006.
373 Supplementing drinking water with Solutein did not mitigate acute morbidity effects of porcine
374 reproductive and respiratory syndrome virus in nursery pigs. *J. Anim. Sci.* 84, 2101–2109.
375 doi:10.2527/jas.2005-616
- 376 Fairbrother, J.M., Nadeau, É., 2019. Colibacillosis, in: Zimmerman, J.J., Karriker, L.A., Ramirez,
377 A., Schwartz, K.J., Stevenson, G.W., Zhang, J. (Eds.), *Diseases of Swine*. John Wiley & Sons,
378 Inc., pp. 807–834.
- 379 Fairbrother, J.M., Nadeau, É., Bélanger, L., Tremblay, C.L., Tremblay, D., Brunelle, M., Wolf, R.,
380 Hellmann, K., Hidalgo, Á., 2017. Immunogenicity and protective efficacy of a single-dose live
381 non-pathogenic *Escherichia coli* oral vaccine against F4-positive enterotoxigenic *Escherichia*
382 *coli* challenge in pigs. *Vaccine* 35, 353–360. doi:10.1016/j.vaccine.2016.11.045
- 383 Fairbrother, J.M., Nadeau, É., Gyles, C.L., 2005. *Escherichia coli* in postweaning diarrhea in pigs:
384 an update on bacterial types, pathogenesis, and prevention strategies . *Anim. Heal. Res. Rev.* 6,

385 17–39. doi:10.1079/ahr2005105

386 García-Meniño, I., García, V., Alonso, M.P., Blanco, J.E., Blanco, J., Mora, A., 2021. Clones of
387 enterotoxigenic and Shiga toxin-producing *Escherichia coli* implicated in swine enteric
388 colibacillosis in Spain and rates of antibiotic resistance. *Vet. Microbiol.* 252.
389 doi:10.1016/j.vetmic.2020.108924

390 Geldhof, M.F., Van Breedam, W., De Jong, E., Lopez Rodriguez, A., Karniychuk, U.U., Vanhee,
391 M., Van Doorselaere, J., Maes, D., Nauwynck, H.J., 2013. Antibody response and maternal
392 immunity upon boosting PRRSV-immune sows with experimental farm-specific and
393 commercial PRRSV vaccines. *Vet. Microbiol.* 167, 260–271.
394 doi:10.1016/j.vetmic.2013.08.017

395 Helm, E.T., Curry, S.M., De Mille, C.M., Schweer, W.P., Burrough, E.R., Gabler, N.K., 2020.
396 Impact of viral disease hypophagia on pig jejunal function and integrity. *PLoS One* 15, 1–18.
397 doi:10.1371/journal.pone.0227265

398 Hsueh, F.C., Wang, S.Y., Lin, W.H., Lin, C.F., Tsai, C.Y., Huang, C.W., Sun, N., Chiou, M.T., Lin,
399 C.N., 2021. Correlation of neutralizing antibodies (Nabs) between sows and piglets and
400 evaluation of protectability associated with maternally derived nabs in pigs against circulating
401 porcine reproductive and respiratory syndrome virus (prrsv) under field conditions. *Vaccines*
402 9. doi:10.3390/vaccines9050414

403 Jha, R., Berrococo, J.F.D., 2016. Dietary fiber and protein fermentation in the intestine of swine and
404 their interactive effects on gut health and on the environment: A review. *Anim. Feed Sci.*
405 *Technol.* 212, 18–26. doi:10.1016/j.anifeedsci.2015.12.002

406 Kitikoon, P., Vincent, A.L., Jones, K.R., Nilubol, D., Yu, S., Janke, B.H., Thacker, B.J., Thacker,
407 E.L., 2009. Vaccine efficacy and immune response to swine influenza virus challenge in pigs
408 infected with porcine reproductive and respiratory syndrome virus at the time of SIV
409 vaccination. *Vet. Microbiol.* 139, 235–244. doi:10.1016/j.vetmic.2009.06.003

410 Lee, S.A., Febery, E., Wilcock, P., Bedford, M.R., 2021. Application of creep feed and phytase
411 super-dosing as tools to support digestive adaption and feed efficiency in piglets at weaning.
412 *Animals* 11. doi:10.3390/ani11072080

413 Lopez, O.J., Oliveira, M.F., Alvarez Garcia, E., Kwon, B.J., Doster, A., Osorio, F.A., 2007.
414 Protection against Porcine Reproductive and Respiratory Syndrome Virus (PRRSV) infection
415 through passive transfer of PRRSV-neutralizing antibodies is dose dependent. *Clin. Vaccine*
416 *Immunol.* 14, 269–275. doi:10.1128/CVI.00304-06

417 Lunney, J.K., Fang, Y., Ladinig, A., Chen, N., Li, Y., Rowland, B., Renukaradhya, G.J., 2016.
418 Porcine reproductive and respiratory syndrome virus (PRRSV): Pathogenesis and interaction
419 with the immune system. *Annu. Rev. Anim. Biosci.* 4, 129–154. doi:10.1146/annurev-animal-
420 022114-111025

421 Luppi, A., 2017. Swine enteric colibacillosis: Diagnosis, therapy and antimicrobial resistance. *Porc.*
422 *Heal. Manag.* 3, 1–18. doi:10.1186/s40813-017-0063-4

423 Martín-Valls, G.E., Mortensen, P., Clilvert, H., Li, Y., Cortey, M., Sno, M., Barna, T., Terré, M.,
424 Guerra, N., Mateu, E., 2022. The use of a whole inactivated PRRS virus vaccine administered
425 in sows and impact on maternally derived immunity and timing of PRRS virus infection in
426 piglets. *Vet. Rec. Open* 9. doi:10.1002/vro2.34

427 McLamb, B.L., Gibson, A.J., Overman, E.L., Stahl, C., Moeser, A.J., 2013. Early Weaning Stress
428 in Pigs Impairs Innate Mucosal Immune Responses to Enterotoxigenic *E. coli* Challenge and
429 Exacerbates Intestinal Injury and Clinical Disease. *PLoS One* 8, 1–12.
430 doi:10.1371/journal.pone.0059838

431 Melkebeek, V., Goddeeris, B.M., Cox, E., 2013. ETEC vaccination in pigs. *Vet. Immunol.*
432 *Immunopathol.* 152, 37–42. doi:10.1016/j.vetimm.2012.09.024

433 Ming, D., Wang, W., Huang, C., Wang, Z., Shi, C., Ding, J., Liu, H., Wang, F., 2021. Effects of
434 Weaning Age at 21 and 28 Days on Growth Performance, Intestinal Morphology and Redox

435 Status in Piglets. *Animals* 11, 1–12. doi:10.3390/ani11082169

436 Nadeau, Fairbrother, J.M., Zentek, J., Bélanger, L., Tremblay, D., Tremblay, C.L., Röhe, I., Vahjen,
437 W., Brunelle, M., Hellmann, K., Cvejić, D., Brunner, B., Schneider, C., Bauer, K., Wolf, R.,
438 Hidalgo, 2017. Efficacy of a single oral dose of a live bivalent *E. coli* vaccine against post-
439 weaning diarrhea due to F4 and F18-positive enterotoxigenic *E. coli*. *Vet. J.* 226, 32–39.
440 doi:10.1016/j.tvjl.2017.07.004

441 Progressis, 2020. . Agencia Española Medicamentos y Productos Sanitarios. URL
442 https://cimavet.aemps.es/cimavet/pdfs/es/ft/1359+ESP/FT_1359+ESP.pdf (accessed 10.14.22).

443 R Core Team. R: A Language and Environment for Statistical Computing, Vienna, Austria.
444 Available online: <http://www.R-project.org/> (accessed on 10.14.22).

445 Ramis, G., Pérez-Esteruelas, L., Gómez-Cabrera, C.G., de Pascual-Monreal, C., Gonzalez-Guijarro,
446 B., Párraga-Ros, E., Sánchez-Uribe, P., Claver-Mateos, M., Mendonça-Pascoal, L., Martínez-
447 Alarcón, L., 2022. Oral and Parenteral Vaccination against *Escherichia coli* in Piglets Results
448 in Different Responses. *Animals* 12. doi:10.3390/ani12202758

449 Rhouma, M., Fairbrother, J.M., Beaudry, F., Letellier, A., 2017. Post weaning diarrhea in pigs: Risk
450 factors and non-colistin-based control strategies. *Acta Vet. Scand.* 59, 1–19.
451 doi:10.1186/s13028-017-0299-7

452 Sang, Y., Rowland, R.R.R., Blecha, F., 2011. Interaction between innate immunity and porcine
453 reproductive and respiratory syndrome virus. *Anim. Heal. Res. Rev.* 12, 149–167.
454 doi:10.1017/S1466252311000144

455 Segalés, J., Domingo, M., Chianini, F., Majó, N., Domínguez, J., Darwich, L., Mateu, E., 2004.
456 Immunosuppression in postweaning multisystemic wasting syndrome affected pigs. *Vet.*
457 *Microbiol.* 98, 151–158. doi:10.1016/j.vetmic.2003.10.007

458 The European Medicines Agency - EMA, 2017. Questions and answers on veterinary medicinal
459 products containing zinc oxide to be administered orally to food producing species. Outcome

460 of a referral procedure under article 35 of Directive 2001/82/EC (EMEA/V/A/118). URL
461 [https://www.ema.europa.eu/en/documents/referral/zinc-oxide-article-35-referral-questions-](https://www.ema.europa.eu/en/documents/referral/zinc-oxide-article-35-referral-questions-answers-veterinary-medicinal-products-containing-zinc-oxide_en.pdf)
462 [answers-veterinary-medicinal-products-containing-zinc-oxide_en.pdf](https://www.ema.europa.eu/en/documents/referral/zinc-oxide-article-35-referral-questions-answers-veterinary-medicinal-products-containing-zinc-oxide_en.pdf) (accessed 10.14.22).

463 Thomson, J.R., Friendship, R.M., 2019. Digestive System, in: Zimmerman, J.J., Karriker, L.A.,
464 Ramirez, A., Schwartz, K.J., Stevenson, G.W., Zhang, J. (Eds.), Diseases of Swine. Wiley-
465 Blackwell, pp. 234–263.

466 Van der Stede, Y., Cox, E., Verdonck, F., Vancaeneghem, S., Goddeeris, B.M., 2003. Reduced
467 faecal excretion of F4+ *E. coli* by the intramuscular immunisation of suckling piglets by the
468 addition of 1,25-dihydroxyvitamin D3 or CpG-oligodeoxynucleotides. *Vaccine* 21, 1023–
469 1032.

470 Van der Weken, H., Sanz Garcia, R., Sanders, N.N., Cox, E., Devriendt, B., 2021. Antibody-
471 Mediated Targeting of Antigens to Intestinal Aminopeptidase N Elicits Gut IgA Responses in
472 Pigs. *Front. Immunol.* 12, 1–15. doi:10.3389/fimmu.2021.753371

473 Vangroenweghe, F., Thas, O., 2020. Improved Piglet Performance and Reduced Antibiotic Use
474 Following Oral Vaccination with a Live Avirulent *Escherichia Coli* F4 Vaccine against Post-
475 Weaning Diarrhea. *J. Clin. Res. Med.* 3, 1–8. doi:10.31038/jcrm.2020309

476 Vangroenweghe, F.A.C.J., Boone, M., 2022. Vaccination with an *Escherichia coli* F4/F18 Vaccine
477 Improves Piglet Performance Combined with a Reduction in Antimicrobial Use and Secondary
478 Infections Due to *Streptococcus suis*. *Animals* 12. doi:10.3390/ani12172231

479 Zentek, J., Buchheit-Renko, S., Männer, K., Pieper, R., Vahjen, W., 2012. Intestinal concentrations
480 of free and encapsulated dietary medium-chain fatty acids and effects on gastric microbial
481 ecology and bacterial metabolic products in the digestive tract of piglets. *Arch. Anim. Nutr.*
482 66, 14–26. doi:10.1080/1745039X.2011.644916

483 Zhao, J., Wan, S., Sun, N., Sun, P., Sun, Y., Khan, A., Guo, J., Zheng, X., Fan, K., Yin, W., Li, H.,
484 2021. Damage to intestinal barrier integrity in piglets caused by porcine reproductive and

485 respiratory syndrome virus infection. *Vet. Res.* 52, 93. doi:10.1186/s13567-021-00965-3

486

487 **Figure captions**

488 **Figure 1.** Average of piglet weight (expressed as mean \pm standard error mean) for piglets in each
489 group and age. Different letters indicate significant differences among groups at each age.

490

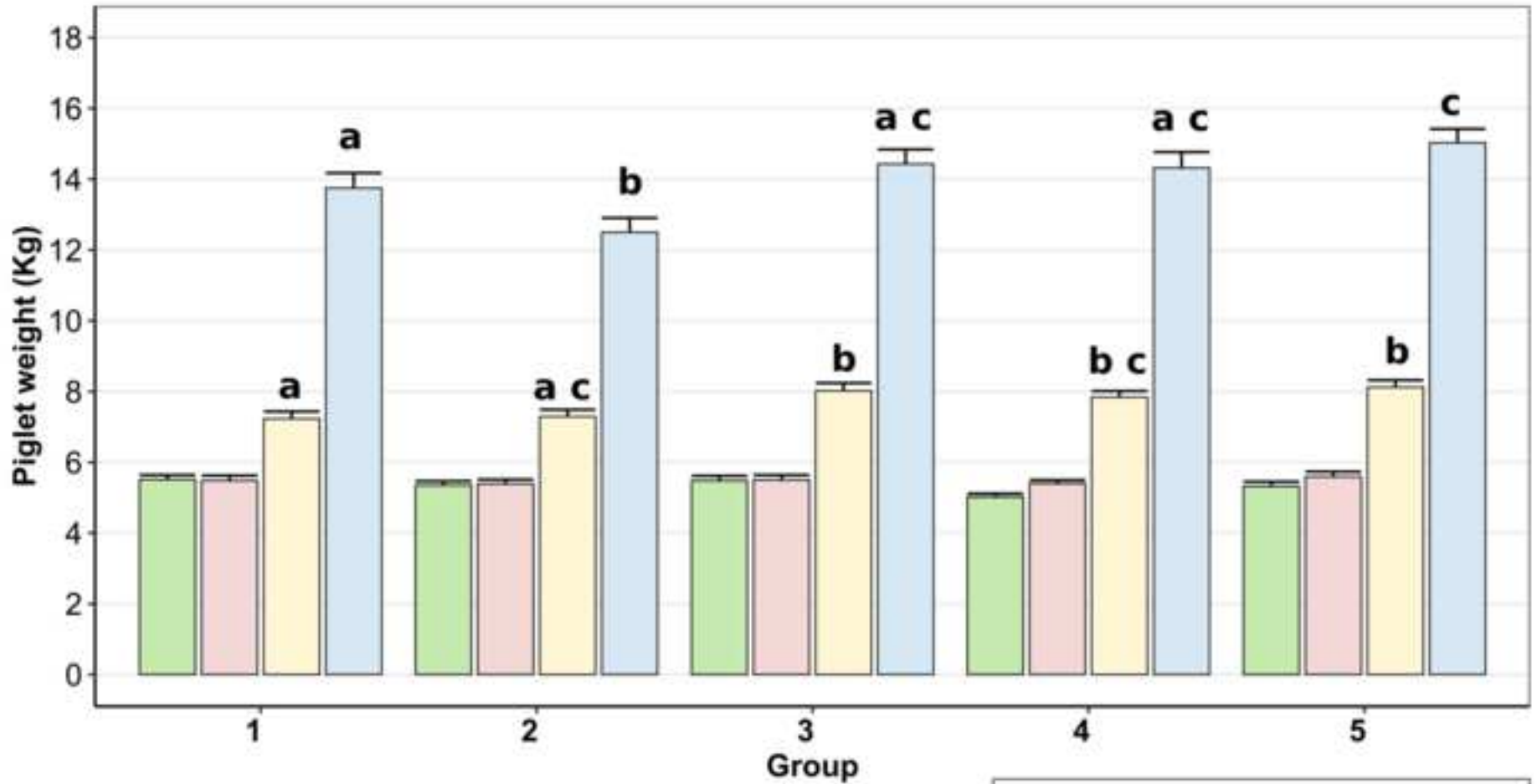
491 **Figure 2.** Average of piglet ADWG (expressed as mean \pm standard error mean) for piglets in each
492 group and feeding period. Different letters indicate significant differences among groups at each
493 feeding period.

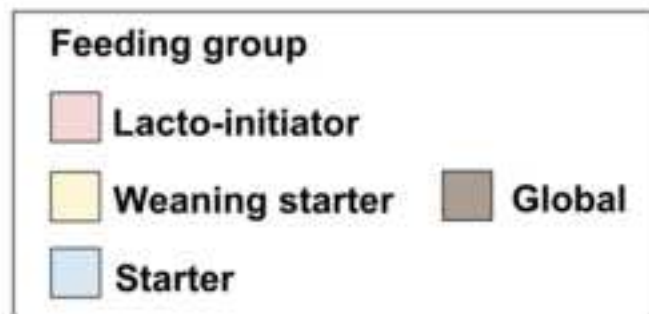
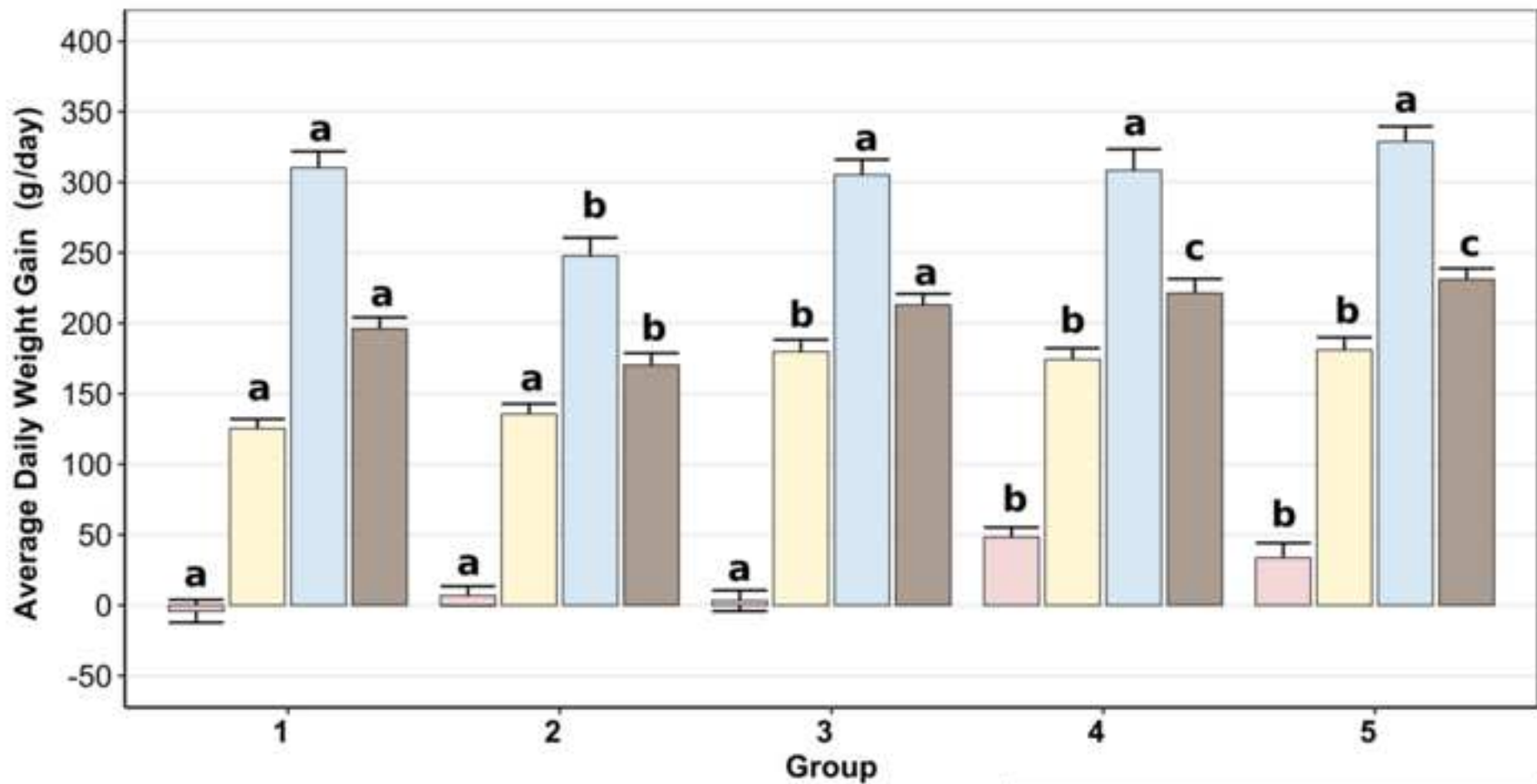
494

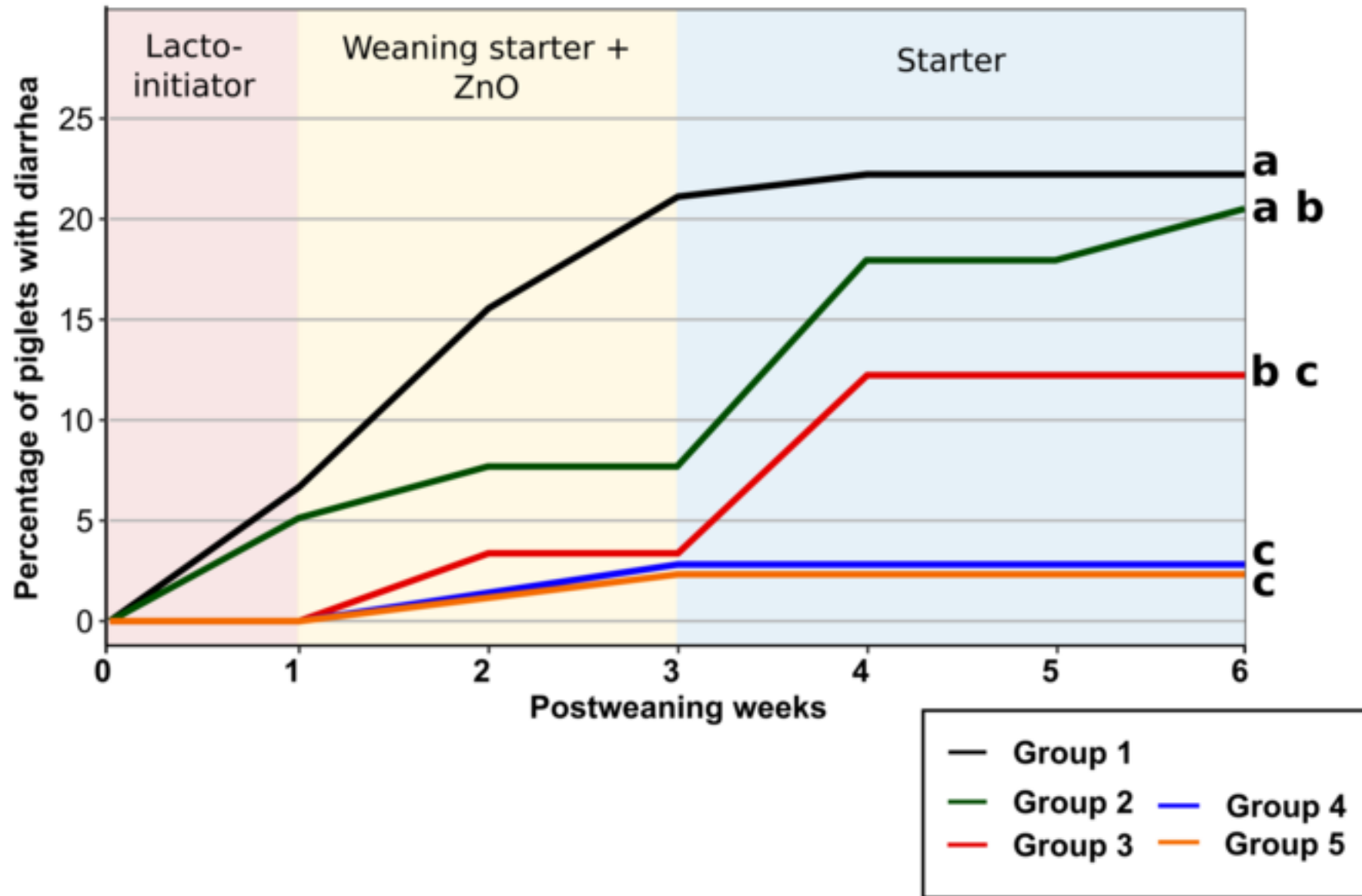
495 **Figure 3.** Evolution of the percentage of piglets with diarrhea in each group. Different letters
496 indicate significant differences in the final proportion of piglets with diarrhea.

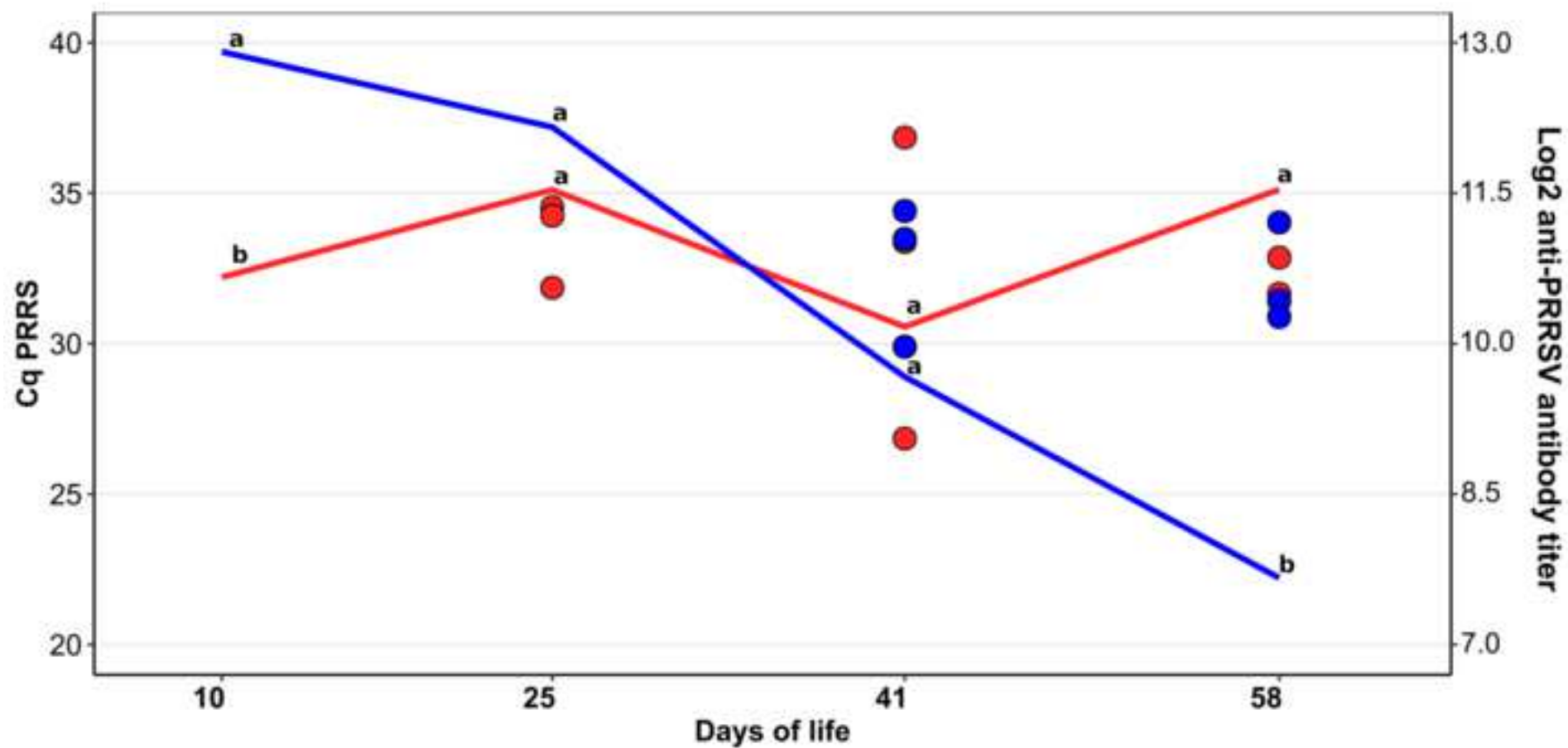
497

498 **Figure 4.** Dynamic of PRRS antibody titer and PRRS viremia in piglets from unvaccinated and
499 vaccinated sows. The lines represent the evolution of the mean of PRRS antibody titer (individual
500 samples) and the dots indicate the Cq value obtained in the positive pools. Different letters indicate
501 significant differences in the PRRS antibody titer in each sampling age.









● Piglets from PRRS unvaccinated sows ● Piglets from PRRS vaccinated sows

Table 1. Groups included in the study and *E. coli* and PRRSV vaccination programs.

Group	Pathogen	Route	Product	Schedule
Group 1 – Control (n = 90).	<i>E. coli</i>	Piglets not vaccinated		
	PRRSV	Piglets from PRRS non-vaccinated sows		
Group 2 – IM <i>E. coli</i> vaccination (n = 78)	<i>E. coli</i>	Intramuscular	Colidex-C ¹	0.5 and 1 mL at 10 and 21 days old, respectively
	PRRSV	Piglets from PRRS non-vaccinated sows		
Group 3 – oral <i>E. coli</i> vaccination (n = 89).	<i>E. coli</i>	Oral	Coliprotec F4/F18 ²	One dose: 2 mL at 21 days old
	PRRSV	Piglets from PRRS non-vaccinated sows		
Group 4 – IM <i>E. coli</i> vaccination + PRRSV vaccination (n = 71).	<i>E. coli</i>	Intramuscular	Colidex-C ¹	0.5 and 1 mL at 10 and 21 days old, respectively
	PRRSV	Piglets from PRRS vaccinated sows	Progressis ³	2 mL at 60 and 21 days before farrowing, respectively
Group 5 – oral <i>E. coli</i> vaccination + PRRSV vaccination (n = 86).	<i>E. coli</i>	Oral	Coliprotec F4/F18 ²	One dose: 2 mL at 21 days old
	PRRSV	Piglets from PRRS vaccinated sows	Progressis ³	2 mL at 60 and 21 days before farrowing, respectively

¹ Vetia Animal Health, San Sebastián de los Reyes, Spain.

² Elanco GmbH, Heinz-Lohman-Str, Germany.

³ Ceva Salud Animal, Barcelona, Spain.

Table 2. Food intake and FCR for each phase of the feeding strategy.

Group	Lacto-initiator		Weaning starter		Starter	
	Food intake (kg/piglet)	FCR (kg food)	Food intake (kg/piglet)	FCR (kg food)	Food intake (kg/piglet)	FCR (kg food)
Group 1	0.489	-5.15	2.72	1.80	10.40	1.65
Group 2	0.421	131.4	3.11	1.77	9.06	1.84
Group 3	0.532	150.2	3.66	1.56	11.44	1.78
Group 4	0.903	2.56	3.76	1.60	11.00	1.76
Group 5	0.674	2.66	4.08	1.83	12.18	1.87

Table 3. Postweaning diarrhea outcomes and percentage of accumulated mortality

Group	Piglets with clinical diarrhea/Total piglets (%)	Days with clinical diarrhea ($\bar{X} \pm sd$)	Total antibiotic injections to treat diarrhea (injections per piglet with clinical diarrhea)	Dead piglets affected by diarrhea/total dead piglets (%)	Dead piglets/total piglets with diarrhea (%)	Dead piglets /total piglets (% accumulated mortality)
Group 1	20/90 (22.22) ^a	2.53 ± 1.50 ^a	50 (2.50)	7/13 (53.85)	7/20 (35.00)	13/90 (14.44) ^a
Group 2	16/78 (20.51) ^{a,b}	1.63 ± 0.72 ^b	26 (1.63)	2/5 (40.00)	2/16 (12.50)	5/78 (6.41) ^{a,b}
Group 3	9/89 (10.11) ^{b,c}	1.56 ± 0.53 ^b	14 (1.56)	2/7 (28.57)	2/9 (22.22)	7/89 (7.86) ^{a,b}
Group 4	2/71 (2.81) ^c	1.50 ± 0.71 ^{a,b}	3 (1.50)	2/3 (66.67)	2/2 (100)	3/71 (4.23) ^b
Group 5	2/86 (2.33) ^c	1.50 ± 0.71 ^{a,b}	3 (1.50)	1/11 (9.09)	1/2 (50.00)	11/86 (12.79) ^{a,b}

^a Different superscript letters indicate statistically significant differences ($p < 0.05$)