

Research priorities for freshwater mussel conservation assessment

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1 **Research priorities for freshwater mussel conservation assessment**

2 **Abstract**

3 Freshwater mussels are declining globally, and effective conservation requires prioritizing research
4 and actions to identify and mitigate threats impacting mussel species. Conservation priorities vary
5 widely, ranging from preventing imminent extinction to maintaining abundant populations. Here, we
6 develop a portfolio of priority research topics for freshwater mussel conservation assessment. To
7 address these topics, we group research priorities into two categories, intrinsic or extrinsic factors.
8 Intrinsic factors are indicators of organismal or population status, while extrinsic factors encompass
9 environmental variables and threats. An understanding of intrinsic factors is useful in monitoring,
10 and of extrinsic factors are important to understand ongoing and potential impacts on conservation
11 status. This dual approach can guide conservation status assessments prior to the establishment of
12 priority species and implementation of conservation management actions.

13 **Keywords:** Unionida; Intrinsic factors; Extrinsic factors; Indicators; Threats; Management.

14 **1. Introduction**

15 Freshwater mussels (Mollusca: Bivalvia: Unionida) are benthic macroinvertebrates that use their
16 muscular foot and shell to burrow into the sediment (Allen and Vaughn, 2009). Adults filter-feed on
17 particles from the water column and interstitial space using cilia-generated water currents (Vaughn
18 et al., 2008; Walker et al., 2014). They have a unique life cycle in which the larva (glochidia, lasidia,
19 or haustoria) must attach to a vertebrate host, usually a fish, and subsequently metamorphose into
20 a juvenile mussel (Wächtler et al., 2001).

21 Scientific interest in freshwater mussels has grown dramatically since the 1970s (Strayer et al., 2004;
22 Lopes-Lima et al., 2014) when the first modern extinctions were recognized (Stansberry, 1970;
23 1971). As currently defined, the Order Unionida, the freshwater mussels, comprises six nominal
24 families and around 800 described species, although the exact number fluctuates as new species are
25 described and taxonomic revisions to existing taxa are made (Williams et al., 2017; Graf and
26 Cummings, 2018). In this context, accurate taxonomic identification plays a key role in species
27 conservation, and modern phylogenetic information is a critical component of conservation biology
28 (Morrison et al., 2009). Freshwater mussels are globally imperiled (6% of known species having
29 recently become extinct; IUCN, 2017), with declines in distribution and abundance related to a
30 variety of factors including habitat modification, water quality degradation, climate change,
31 introduction of non-native species, declines in fish hosts, and over-exploitation (Strayer et al., 2004;
32 Walker et al., 2014; Lopes-Lima et al., 2018). Of 535 freshwater mussel species assessed by the

33 International Union for Conservation of Nature (IUCN, 2017), 217 were categorized as Near
34 Threatened, Vulnerable, Endangered, or Critically Endangered, and 89 species were classified as Data
35 Deficient (Fig. 1). Given these high levels of imperilment, establishing research priorities that support
36 more accurate determination of a species' status is critical. Here we summarize the most important
37 research needs for assessing the conservation status of freshwater mussels (Table 1) and discuss
38 how practitioners can leverage this information to improve the development and implementation of
39 effective conservation and management strategies.

40 Characters used to assess the conservation status of a given group of organisms can be subdivided
41 into intrinsic and extrinsic factors (following Williams et al., 2008; Dawson et al., 2011; Fig. 2).
42 Intrinsic factors are measures of a species' condition and provide valuable information about
43 demographic trends (e.g., abundance, distribution, and viability of extant populations) and
44 population health status (e.g., reproduction, nutritional status, genetic variability, and growth).
45 Extrinsic factors describe the state of the environment and include factors such as habitat condition
46 and availability, water quality and quantity, impacts of invasive species, and flow alterations, among
47 other ecologically-mediated threats.

48 Individuals respond to extrinsic factors through phenotypic plasticity, and populations respond
49 through adaptation (Fisher, 1930). Phenotypic plasticity has limits, and the potential for
50 evolutionary adaptation depends on genetic polymorphism within populations. Thus, for
51 conservation of at-risk species, it is important to identify which intrinsic factors make these species
52 vulnerable to environmental change (Fig. 3) to help guide conservation actions in a changing
53 environment (e.g., habitat restoration). This includes adaptive genetic variation, which represents
54 the basis for evolutionary change in the future, but is largely unexplored for freshwater mussels.

55 In this paper, we assess the research needed to guide conservation status assessments and future
56 conservation actions for freshwater mussels by summarizing the knowledge of a multidisciplinary,
57 global team of scientists working in this area. We divide research priorities into two categories. First,
58 we discuss how an understanding of intrinsic factors can help us better assess changes in freshwater
59 mussel populations and species over space and time. Second, we identify extrinsic factors that can
60 influence such changes. Lastly, we discuss some strategies to guide freshwater mussel protection,
61 conservation and management.

62 **2. Intrinsic factors**

63 2.1. Species identity

64 Because actual conservation and policies and actions generally occur at the species level, accurate
65 species identification and delimitation is critical. Historically, most taxonomic work has focused on
66 Europe and North America, and even there some species are still being revised and new species are
67 being described (Froufe et al., 2017; Williams et al., 2017; Smith et al., 2018; Pieri et al., 2018).
68 However, mussel communities in Africa, Asia, Australasia, South and Central America have been
69 understudied and merit much more attention (Walker et al., 2014; Zieritz et al., 2018). For example,
70 almost every large or medium-sized tropical freshwater basin in Indo-China is a separate
71 evolutionary hotspot harboring a unique endemic freshwater mussel fauna, and several new genera
72 and species were recently described from this region (Bolotov et al., 2017a, 2017b). However,
73 taxonomic revision of mussel species-level classification is complicated by centuries of “over-
74 description”. For example, there are more than 400 synonyms for *Anodonta cygnea* (Graf and
75 Cummings, 2018). Further, the characters used to define species have been a persistent and
76 contentious problem. Conchological characters traditionally used to identify taxonomic units can be
77 phenotypically plastic and dependent on ecological conditions (Zieritz et al., 2010; Sheldon, 2017). In
78 contrast, studies using molecular data and assessing genetic variability have suggested cryptic
79 speciation where there is no clear morphological differentiation (e.g., Baker et al., 2003; Prié and
80 Puillandre, 2014; Graf et al., 2015; Sheldon, 2017; Araujo et al., 2018; Pieri et al., 2018).

81 Species identity is often incompletely delimited and is a fundamental conservation priority.
82 Morphology combined with molecular phylogenetic information focused on taxa of conservation
83 concern can dramatically improve this situation (Smith et al., 2018). Application of mitochondrial
84 and nuclear DNA markers, as well as next-generation sequencing technologies, and the application
85 of phylogenetic species delimitation methods are promising approaches to improve our
86 understanding of freshwater mussel diversity (Inoue et al., 2018; Smith et al., 2018).

87 2.2. Population size and species distribution

88 Rare or endangered species face genetic risks from small population or range sizes (e.g., genetic
89 drift, inbreeding depression, self-fertilization, and low gene flow; Geist and Kuehn, 2005). In
90 freshwater mussel conservation, much emphasis has been placed on the local abundance of a
91 species, and little on the overall abundance of that species across its entire range and connections
92 between local populations in a larger, regional metapopulation (Newton et al., 2008). Our
93 knowledge about the spatial delimitation of metapopulations is scarce (Terui et al., 2014). Moreover,
94 minimum viable population sizes vary among species, but there are no globally objective criteria
95 used to establish a baseline (Nunney and Campbell, 1993) and there is little knowledge about which
96 abundances are viable for most species. However, against this background it should be recognized

97 that some species have always been rare or have always occupied a limited geographic area. In this
98 regard, low abundance or a small distribution *per se* does not necessarily indicate imperilment
99 (Geist, 2010; Geist et al., 2010). Nevertheless, species with small distributions are more vulnerable
100 to localized human activities, and to environmental and demographic stochasticity.

101 Assessing population abundance based solely on field observations is costly and time-consuming,
102 but can be done by trained volunteers or scientists. Molecular techniques offer options for
103 complementing field-based observations, but should not be viewed as alternatives. Rather,
104 molecular techniques can provide information complementary to field observations. Such molecular
105 techniques (e.g., microsatellites; Froufe et al., 2016) require minimal effort to estimate many
106 biological parameters of interest, including demographic parameters (e.g., abundance, occupancy,
107 disease status), population genetic parameters (e.g., genetic diversity, structure, effective
108 population size), and responses to selective pressures (e.g., exploitation, climate change) (reviewed
109 by Schwartz et al., 2007). In addition, it is possible to use molecular markers to monitor and estimate
110 the level of migration, invasion, and hybridization in wild populations (Koneva, 2013). The
111 development of noninvasive sampling using environmental DNA (eDNA) is also promising, especially
112 for detecting the presence of species under difficult sampling conditions (Sansom and Sassoubre,
113 2017).

114 2.3. Species trends

115 Trends in abundance or distribution are a central consideration in assigning a threat rankings to
116 species. Species trends can be revealed by comparing current and historical data on population sizes
117 and distributions (Bolotov et al., 2012). For example, in France, species distribution models were
118 used to infer range contraction, comparing the extent of occurrence predicted by the models to the
119 currently known extent of occurrence of seven freshwater mussel species (Prié et al., 2014). Popejoy
120 et al. (2018) used zooarchaeological data to characterize freshwater mussel assemblage composition
121 before European colonization of the American continent and compare it to recent mussel
122 communities. Similarly, in Australia, initial attempts have been made to compare the species
123 composition of Aboriginal middens with current populations to understand changes in species
124 distributions (Garvey, 2017). Information from historic and contemporary museum collections and
125 field observations can be used to determine spatiotemporal changes in species' ranges (e.g.,
126 Johnson et al., 2016; Randklev et al., 2018). Interviews with local people (i.e., "citizen scientists")
127 also could provide important information about changes in the distribution and abundance of
128 freshwater mussels over time.

129 Understanding the historic range of a species is critical in determining its temporal trend; however,
130 estimating trends with precision can be difficult. The IUCN approach for determining the Green and
131 Red Lists of threatened species is to examine changes in population size, areas of occupancy and
132 changes in extents of occurrence over set time periods. These estimations also can be provided by
133 using molecular methods such as Bayesian Skyline Plots (i.e., studies of DNA to estimate the
134 genealogy and the relative timing and duration of past demographic events; Drummond et al.,
135 2005), which have been rarely used in demographic studies of freshwater mussels (but see Jones et
136 al., 2015).

137 2.4. Demography

138 Absence of recruitment is a major process driving freshwater mussel declines (e.g., Strayer and
139 Malcom, 2012). The age structure of a mussel population reflects past recruitment events and can
140 be used to project future population trends using conventional demographic modeling tools (e.g.,
141 life tables; Vandermeer and Goldberg, 2003). Except for extremely long-lived species, a prevalence
142 of adult mussels with limited or no younger cohorts may mean that recruitment has declined or
143 failed and that the species is under extinction threat (i.e., “extinction debt”; Haag, 2012).

144 Some long-lived mussel species can persist with infrequent and low recruitment events, even though
145 juvenile survival can be quite low (e.g., *Margaritifera auricularia*; Prié et al., 2018). Other species
146 require more frequent and even annual recruitment to maintain sustainable populations (Outeiro et
147 al., 2008; Hastie et al., 2010; Klunzinger et al., 2014). Factors leading to low recruitment include
148 juvenile habitat degradation and decreases in the populations of fish hosts (Bolotov et al., 2012).

149 Demographic studies examining reproduction, recruitment, population age structure, sex ratios,
150 mortality rates, immigration, and emigration within a metapopulation are of importance for early
151 detection of decline, especially in threatened species (Haag, 2012). Determining demographic
152 structure requires adequate sampling strategies (e.g., Lois et al., 2014) to locate different juvenile
153 stages and determine spawning and recruitment patterns – seasonal or aseasonal.

154 2.5. Life-history traits

155 Many unionid species are long-lived (e.g., *M. margaritifera* can live up to 280 years; Lopes-Lima et
156 al., 2017), with slow growth, late sexual maturity, and low recruitment compared to other groups of
157 freshwater invertebrates (Haag, 2012). However, these life-history parameters can be highly variable
158 within and among species. In this regard, reproductive timing is highly dependent on climate and
159 other environmental factors. For instance, temperature influences the length of the larval
160 development period, and local hydrology can determine larval release (e.g., Hughes et al., 2004;

161 Strayer, 2008; Araujo and Álvarez-Cobelas, 2016; Bunn et al., 2006). An understanding of
162 reproductive phenology and other life history traits can be used to predict local colonization,
163 extinction patterns and provide insights into the long-term persistence of populations (Vaughn,
164 2012).

165 Reproductive traits that might be included as research priorities for assessing mussel conservation
166 status include strategies for attracting fish hosts and releasing larvae (e.g., broadcast of free larvae,
167 conglomerates, and mantle displays; Barnhart et al., 2008), the timing and length of different
168 reproductive stages (gamete production, larval brooding, larval release), fecundity, lifespan, age at
169 sexual maturity, and generation length (average age of parents of the current cohort). With this aim,
170 non-destructive sampling methods such as monitoring free oocytes in ovarian follicles (Dudgeon and
171 Morton, 1983) or examining marsupial brood development in live individuals (Reid et al., 2012) or
172 examination of host fishes for mussel glochidia (Salonen and Taskinen, 2017) are recommended,
173 especially in rare, endangered and long-lived species (Saha and Layzer, 2008).

174 **3. Extrinsic factors**

175 3.1. Fish hosts

176 Conservation of freshwater mussels requires understanding the status of their fish hosts and
177 conserving them as well. Most unionid mussels have obligate ectoparasitic larvae which attach to
178 the gills, fins or other body surfaces of their hosts (for exceptions, see Modesto et al., 2018).
179 Accordingly, mussels must co-occur with appropriate vertebrate hosts to complete their life cycle
180 and allow long-term survival of the population. In addition, movement of infested fish hosts is
181 important to maintain connectivity among mussel metapopulations and upstream dispersal (Zajac et
182 al., 2018).

183 Factors related to larval dispersal ability, host attachment mode, and whether a mussel species is a
184 host generalist or specialist, are key determinants of recruitment success or failure (Newton et al.,
185 2008). In addition, detailed information on parameters such as infestation abundance (infested
186 fish/total number of fish), infestation intensity (larvae/infested fish), host physiological compatibility
187 (metamorphosis success) and host availability (contact probability) is fundamental (e.g., Salonen et
188 al., 2017). This information can be acquired through laboratory experiments to determine fish hosts
189 and subsequent field studies to verify that these relationships occur in nature (e.g., Klunzinger et al.,
190 2012). Sampling fish assemblages to determine natural infestation rates and host population size,
191 distribution and life history can be used to determine host availability and contact probability
192 (Jansen, 1991; Kelly and Watters, 2010). Information on fish host population trends and movement

193 can be used to determine the potential for mussel dispersal, which is important for long-term
194 conservation; e.g., for founding new populations as habitat changes. In addition, many of the factors
195 that affect fish hosts (e.g., habitat degradation, water quality impairment or invasive species) also
196 affect mussel recruitment and can have cascading effects on the entire freshwater mussel
197 assemblage (Geist et al., 2006; Stoeckl et al., 2014).

198 3.2. Habitat characteristics

199 Predicting suitable habitat and the potential distribution of freshwater mussel species is a high
200 priority for conservation (Jones and Byrne, 2014). However, predicting freshwater mussel diversity
201 hotspots differs from assessments for many other aquatic and terrestrial species because it is easier
202 to locate a population than to predict its distribution, and is dependent on the characteristics of the
203 whole catchment (Abell et al., 2006; Graf and Cummings, 2011).

204 An understanding of habitat requirements – leading to development of mussel species distribution
205 models – can be based on field surveys, published information, historical data (including museum
206 collections) on freshwater mussel presence, and local knowledge (Bolotov et al., 2012; Prié et al.,
207 2014). Both natural and anthropogenic factors are important in modeling suitable habitat
208 characteristics to predict species distributions (Mynsberge et al., 2009), including current and
209 historical climates, catchment and stream characteristics, water and sediment characteristics, floral
210 and faunal assemblages, and anthropogenic threats (Strayer, 1999; Howard and Cuffey, 2003; Geist
211 and Auerswald, 2007; Inoue et al., 2015; Klunzinger et al., 2015). However, complete information is
212 rarely available. In addition, in assessing historic sites, recent data on how habitats may have been
213 modified or disturbed should be evaluated to understand which changes led suitable habitats to
214 become unsuitable (e.g., Zieritz et al., 2018). Finally, scale differences and correlations among
215 factors can complicate modelling. The use of high-resolution ecological niche modelling and
216 computer models (e.g., GIS) coupled with field verifications can assist in this effort (Inoue et al.,
217 2015; Walters et al., 2017).

218 3.3. Catchment use

219 Anthropogenic land-cover alteration is one of the biggest drivers of global change and profoundly
220 affects biological diversity and ecosystem health (Dynesius and Nilsson, 1994; Allan, 2004). In
221 general, land-use change (e.g., deforestation, agricultural development, livestock production,
222 mining, and drainage of wetlands) (Fig. 4a) can increase erosion and pollution, and change river
223 discharge regimes (Makhrov et al., 2014; Walker et al., 2014), all likely to have impacts on
224 freshwater mussel populations.

225 Deposited and suspended materials impact mussels in two ways: by clogging mussel gills and
226 thereby interfering with filter feeding, and by filling interstitial spaces in substrates and thereby
227 eliminating important habitats for juveniles (Österling et al., 2010; and references therein). Wicklow
228 et al. (2017) found that riparian forest cover was a strong predictor of healthy populations of the
229 imperiled brook floater, *Alasmidonta varicosa*, whereas low or no forest cover was associated with
230 the most vulnerable populations. In Southeast Asia, declines in the endemic mussel fauna appear to
231 be related to deforestation and agriculture (Bolotov et al., 2014; Zieritz et al., 2018). In contrast,
232 agricultural drainages from paddy fields in Japan positively affect mussels living in rivers and
233 agricultural ditches due to increased food abundance and more suitable water temperatures
234 (Nakano, 2017; Nishio et al., 2017). Hence, assessment of the degree of a species' tolerance to
235 impacts associated with land-use change (e.g., sedimentation, turbidity, nutrient pollution) is of
236 paramount importance for predicting the present and future impacts on freshwater mussel
237 assemblages (Gallardo et al., 2018).

238 3.4. Habitat alteration

239 Altered flow (hydrological) and temperature regimes associated with impoundment are one of the
240 most important threats to freshwater fauna generally (Fig. 4b). Impoundments alter seasonal
241 temperature regimes and create lentic or semi-lentic conditions with higher sedimentation and low
242 oxygen concentrations. Therefore, species composition may shift, with reductions in the abundance
243 of riverine taxa and increases in lentic taxa (Pringle et al., 2000). At the same time, releases from
244 reservoirs can alter flow regimes which may increase the probability of mortality of mussels (e.g., by
245 exposure to thermal variations; Gagnon et al., 2004; Haag and Warren, 2008; Araujo and Álvarez-
246 Cobelas, 2016). Furthermore, hypolimnetic dam discharge may negatively affect gametogenesis and
247 glochidial production (Haag, 2012). Structures such as dams, floodgates and hanging culverts can
248 also block the movement or alter the species assemblages of fish hosts (Tiemann et al., 2004;
249 Winemiller et al., 2016), making them less available to mussels, isolating populations, and impacting
250 mussel reproductive success and contributing to overall assemblage depletion (Smith, 1985;
251 Watters, 1996; Kelner and Sietman, 2000; Vaughn, 2012). In extreme cases, mussels have been
252 completely isolated from their fish hosts, resulting in functional extirpation, such as with Ebonyshell,
253 *Reginaia ebenus*, in the Upper Mississippi River (Kelner and Sietman, 2000).

254 Given the unprecedented boom in construction of hydropower dams in South America, Africa and
255 Asia (Winemiller et al., 2016), studies are needed to assess the effects of such habitat alterations on
256 mussels and their fish hosts. Such studies need to assess both individual, cumulative, and interactive
257 impacts of multiple stressors (e.g., low oxygen, increasing or decreasing temperature, flow velocity).

258 In addition, other hydrological and habitat alterations likely to affect freshwater mussels include
259 sand and gravel extraction, channelization, riverine urbanization, and cattle access to rivers.

260 3.5. Pollution

261 Water and sediment pollution from point sources (e.g., chemical or quarry spills, urban or industrial
262 wastewater discharge) and non-point sources (e.g., runoff from agriculture and roads) influences
263 mussels (Grabarkiewicz and Davis, 2008; Gillis et al., 2017). Unionid mussels are generally
264 underrepresented in toxicity databases used for developing of protective water quality criteria and
265 other guidance. When compared with other freshwater organisms, mussel species were among the
266 species most sensitive to a variety of chemical compounds (Wang et al., 2017), with juvenile life
267 stages particularly sensitive to water and sediment pollution (Augspurger, 2003; Cope et al., 2008;
268 Taskinen et al., 2011; Wang et al., 2017; Kleinhenz, 2018).

269 Most toxicity tests have traditionally used survival as the endpoint (Naimo, 1995), and more
270 attention to sublethal effects on physiology and reproduction is needed. As an example, molecular
271 approaches (i.e., gene expression) recently performed in freshwater mussels allow the identification
272 of differentially expressed genes in response to environmental stressors (Michalak et al., 2017;
273 Ferreira-Rodríguez et al., 2018a). In addition to laboratory tests, we need to understand pollution –
274 including micropollutant – effects on ecological condition and demography. This will require
275 extensive testing to include sediment- and diet-borne contaminant exposures (Cope et al., 2008),
276 multiple stressors, field monitoring, and in situ experiments to establish ecological thresholds and
277 translate these into enforceable water quality regulations.

278 3.6. Climate change

279 Climate change threatens freshwater mussels through higher temperatures, and increased
280 frequency and severity of droughts and floods (Kundzewicz et al., 2008; Fig. 4c). The thermal regime
281 of their aquatic habitat is related to larval metamorphosis success (Taeubert et al., 2014), metabolic
282 rate (Ferreira-Rodríguez and Pardo, 2017), and overall survival (Akiyama and Iwakuma, 2007;
283 Pandolfo et al., 2010). Temperature stress also can affect the sensitivity of mussels to additional
284 stressors, such as hypoxia, water pollution or introduction of non-native species (van Hattum et al.,
285 1993; Ferreira-Rodríguez and Pardo, 2017). Additionally, severe droughts associated with extreme
286 temperatures can result in extremely low flows, with declines in species richness and local
287 extirpations (Gagnon et al., 2004; Haag and Warren, 2008; Sousa et al., in press). In contrast,
288 flooding due to extreme precipitation events can scour mussel beds, disrupt recruitment, or hinder

289 species recovery efforts, such as translocations or reintroductions of endangered species (Strayer,
290 1999; Steuer et al., 2008; Haag, 2012; Stodola et al., 2017).

291 The effects of climate change may be exacerbated by increased anthropogenic water withdrawals
292 and river regulation for human needs (Carter et al., 2014; Prein et al., 2016). Moreover, shortened
293 intervals between successive extreme climatic events may not allow sufficient time for freshwater
294 mussel assemblages to recover demographically, particularly for long-lived, low-recruitment species
295 (Vaughn et al., 2015). A combination of modelling and experimental approaches is needed to predict
296 how these changes will restructure mussel assemblages (Santos et al., 2015; Vaughn et al., 2015;
297 Ferreira-Rodríguez and Pardo, 2017).

298 3.7. Non-native bivalve species

299 Biological invasions are also a major component of human-caused global change (Lockwood et al.,
300 2013). The golden mussel *Limnoperna fortunei* in South America and Japan and zebra mussels
301 *Dreissena* spp. in the Northern Hemisphere modify the nature and complexity of the substrate,
302 abundance of plankton and other available food resources, which may negatively affect other
303 freshwater mussels (Darrigran and Damborenea, 2005; Sousa et al., 2014; Fig. 4d). In addition,
304 mussels byssally-attached to unionid shells may hamper their filter feeding, respiration, locomotion,
305 and reproduction capacity by interfering with valve-movement behavior (Lucy et al., 2014). The
306 Asian clam *Corbicula* spp. potentially competes for food and space with freshwater mussels
307 (Ferreira-Rodríguez et al., 2018b). While reproduction of native freshwater mussel species may be
308 constrained by fish immunity, host availability does not constitute a major limit for the Chinese pond
309 mussel *Sinanodonta woodiana*, a broad generalist species (Douda et al., 2017). In addition, in a
310 global warming context, some non-native species cope much better with unfavorable conditions
311 (e.g., thermal extremes) than native species (Bielen et al., 2016). More work is needed to
312 understand how non-native bivalves interact with and affect native mussels over the long term,
313 including impacts on early life stages, to identify which native species can compete successfully with
314 invaders, and to determine which species will face extinction by the additive effect of non-native
315 competitors and other threats (e.g., climate change, pollution).

316 3.8. Other extrinsic factors

317 Other extrinsic factors influencing mussels include diseases and parasitism, exotic fish introductions,
318 increased predation, and over-exploitation. While these factors are often secondary in importance
319 to habitat modification, land-use change, and water quality impairment can interact, becoming
320 cumulative and locally important. Diseases and parasitism are potentially important, but few studies

321 are available to assess their importance (Taskinen and Valtonen, 1995; Grizzle and Brunner, 2009;
322 Carella et al., 2016). An increase in exotic fishes can impair reproduction of freshwater mussels
323 (Taniguchi et al., 2002; Doua et al., 2014). Predation on mussels is usually size-selective upon
324 smaller individuals, thereby affecting the age structure of populations (Tyrrell and Hornbach, 1998;
325 Zajac, 2014). Finally, freshwater mussel species may face local or even global extinction from human
326 over-exploitation, particularly in Asia (Bolotov et al., 2014; Zieritz et al., 2018; Do et al., 2018; Fig.
327 4e). Hence, the incidence of these and other factors should be addressed when assessing the
328 conservation status of the target species/population.

329 **4. Conservation and recovery plans**

330 The development of conservation strategies for freshwater mussels faces many challenges, including
331 the selection of priority species and populations for conservation, strategic decisions about habitat
332 restoration, captive propagation, and stakeholder and general public involvement (Geist, 2010;
333 Strayer, 2017).

334 Laws and regulations to protect freshwater mussels have been enacted (although large differences
335 exist between countries and continents), but very few protected areas have been established as
336 refugia for endangered freshwater mussels (e.g., the Mudpuppy Conservation Area, Missouri, USA;
337 reaches of the Verdigris, Fall, and Neosho Rivers in Kansas, USA). In the European Union, each
338 member state hosting mussel species listed in Annex II of the Habitat's Directive has been required
339 to protect important populations as Special Areas of Conservation (SAC's; Council Directive
340 92/43/EEC). However, the Habitats Directive was published in 1992 with knowledge from the 1980s,
341 and it has not been updated since then. Therefore, the Directive does not reflect the correct number
342 of threatened species, which leaves many taxa unprotected. In addition, external anthropogenic
343 activities affecting freshwater mussels are also likely to affect population dynamics of fish hosts
344 (Modesto et al., 2018); therefore, it is essential to delimit appropriate spatial scales of conservation
345 units (Abell et al., 2006).

346 The conservation genetic literature describes two types of conservation units, evolutionary
347 significant units and management units (Funk et al., 2012). An evolutionary significant unit (ESU) can
348 be generally defined as a population or group of populations with high genetic and ecological
349 distinctiveness from other units. Management units (MUs) are defined as populations that are
350 demographically independent of one another (i.e., population dynamics depends on local birth and
351 death rates and not on genetically effective migration). These principles are critical for conservation
352 purposes in two respects. First, maintaining different ESUs will maximize evolutionary potential in

353 the face of environmental change (Fraser and Bernatchez, 2001) and second, maintaining multiple
354 MUs will help preserve demographic viability of infraspecific units and genetic diversity (Grobler et
355 al., 2006; Zannatta and Harris, 2013). Although the use of neutral genetic markers is essential to
356 diagnose evolutionary histories, further data (such as analysis of demographic trends and
357 population-level evaluations of host compatibility) can be used to reveal ecologically relevant MUs.

358 To ensure the long-term persistence of species, a conservation strategy may include an approach to
359 achieve ecologically sustainable land-use (protection from extrinsic factor modification; see Fig. 2),
360 such as riparian vegetation buffers around waterways (e.g., Giam et al., 2015). However, caution
361 must be exercised when restoration actions are proposed. For example, removing dams can harm
362 freshwater mussels by releasing pollutants or destabilizing sediments. Fish passageways to allow
363 migration past dams may be helpful in reconnecting upstream and downstream populations of both
364 mussels and host fish (e.g., Benson et al., 2017). However, past designs often have been ineffective,
365 and fish passageways sometimes are used to justify harmful dams (Brown et al., 2013). To prevent
366 additional species losses – when the causes of decline are identified and corrected – researchers
367 have developed methods for hatchery propagation of juvenile mussels to supplement, augment, or
368 restore populations. Propagation programs will be critical for many recovery efforts (Carey et al.,
369 2015). However, mussel propagation requires accurate genetic identification of conservation units
370 prior to release of animals produced in hatcheries to ensure preservation of remaining genetic
371 diversity (Jones et al., 2006; McMurray and Roe, 2017).

372 The involvement of local stakeholders, policy makers, local authorities, and others responsible for
373 water management is a major challenge in achieving successful freshwater mussel conservation
374 (e.g., Linehan, 2007). In addition, collaborative efforts will help to harness public interest and
375 knowledge. Similarly, new information and communication technologies provide methods to
376 improve popular discourse and knowledge portrayed in mass and social media about the importance
377 of freshwater mussels (i.e., providing ecosystem services; Vaughn, 2018), thereby increasing social
378 awareness of their values (Strayer, 2017). Certainly, closer cooperation between scientists is needed
379 (e.g., the Freshwater Mollusk Conservation Society, U.S.A.), as well as initiatives such as adopting
380 international standards for monitoring freshwater mussels (e.g., the *M. margaritifera* CEN Standard;
381 British Standards Institution, 2017). Thanks to the knowledge gained through scientific research,
382 techniques that enable the large-scale production of juveniles (including *in vitro* larval
383 metamorphosis without fish hosts) are being developed and applied in recovery programs (e.g.,
384 Patterson et al., 2018). Finally, the quantifiable – temporal and spatial – measurements that we have
385 discussed here provide a synopsis of the conditions and trends of freshwater mussel populations and

386 their habitats. However, without appropriate support and funding, effective conservation will be
387 difficult to implement.

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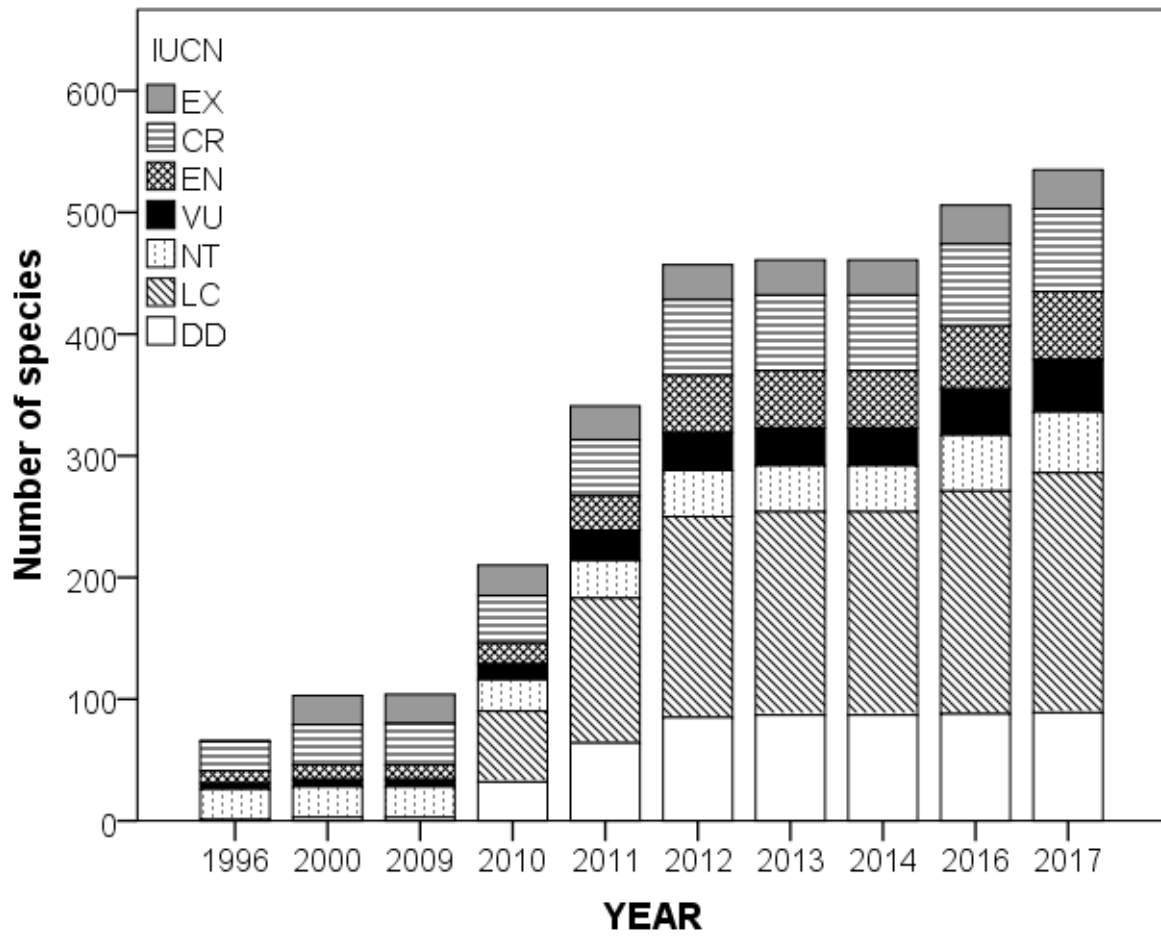
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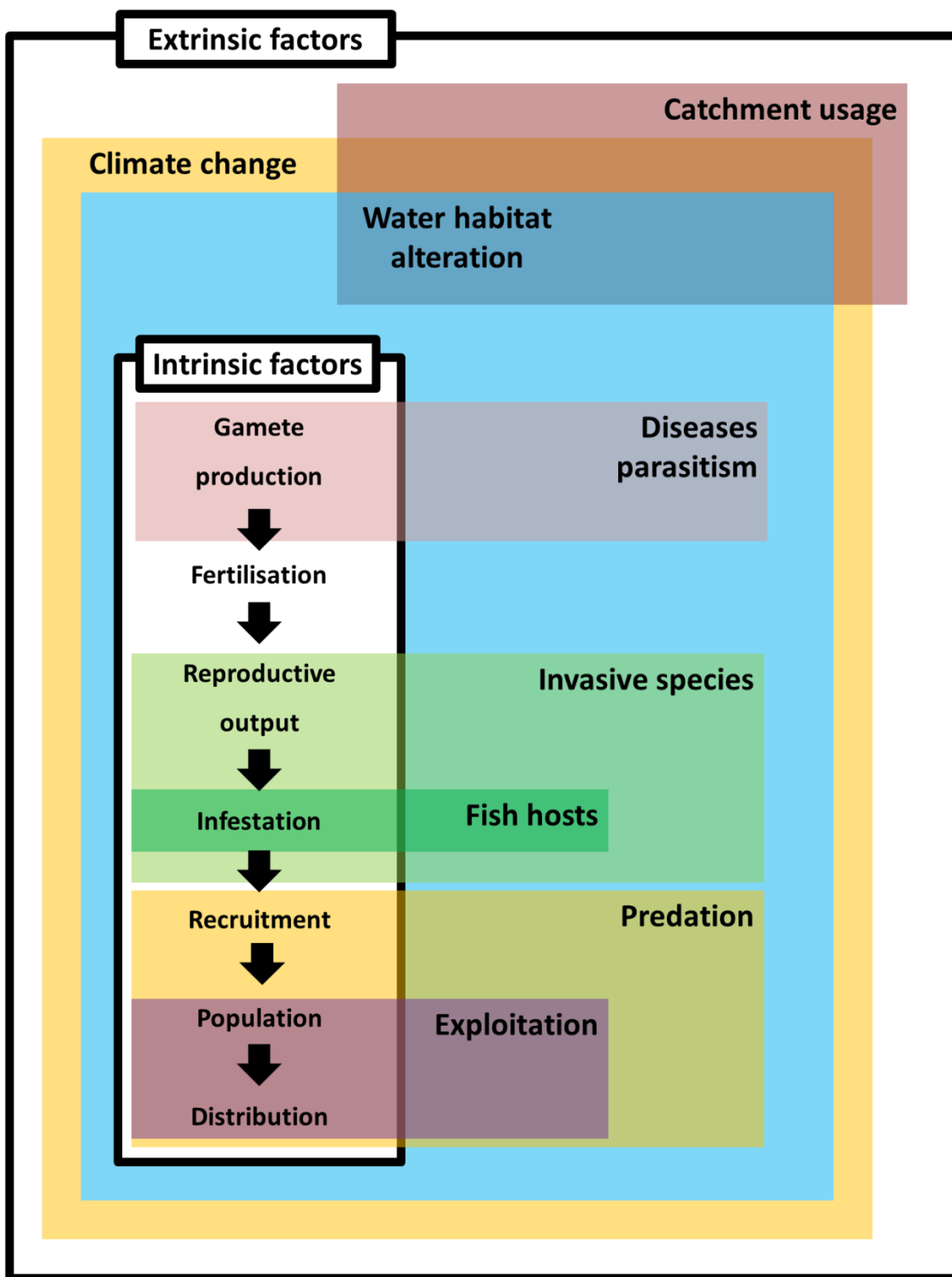
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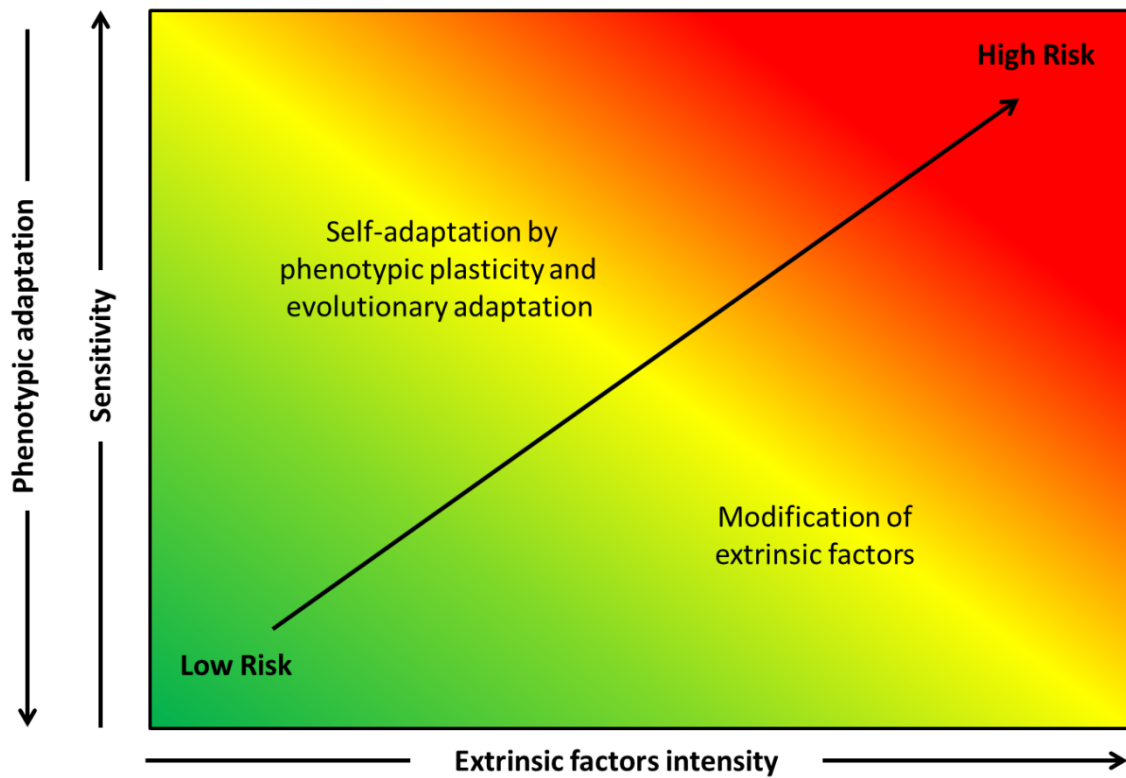
826 **Fig. 1.** Number of assessed freshwater mussel taxa and distribution of IUCN Red list status in recent years. DD, data
 827 deficient; LC, least concern; VU, vulnerable; NT, near threatened; EN, endangered; CR, critically endangered; EX, extinct.

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830 **Fig. 2.** General framework for assessing freshwater mussel conservation status. Intrinsic factors are measures of the
 831 species condition based on individual- and/or population-level processes. Extrinsic factors describe the state of the
 832 environment. All elements of this framework need to be considered in a comprehensive species-level risk assessment.



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834 **Fig. 3.** Species risk assessment based on exposure to extrinsic factor and intrinsic sensitivity. The balance between the
 835 species intrinsic factors able to adapt to environmental change and extrinsic factor modifications determine the
 836 implementation of conservation and/or recovery plans. Adapted from Dawson et al. (2011).

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855 **Fig. 4.** Extrinsic factors responsible for mussel declines. a) Catchment use - agriculture. Stream in Kesang River Basin
 856 (Peninsular Malaysia) in an oil palm plantation. b) Habitat alteration. Dam on the River Miño, Spain-Portugal. c) Climate
 857 change. The North African (Morocco) river Oued Martil in the driest season. d) Non-native *Dreissena polymorpha* byssally-
 858 attached to *Unio pictorum*. e) Harvest. Freshwater mussel collection by local people in the Tauk Ue Kupt River (Sittaung
 859 River basin), Myanmar. f) habitat destruction by cattle, Gingin Brook, southwest Australia.

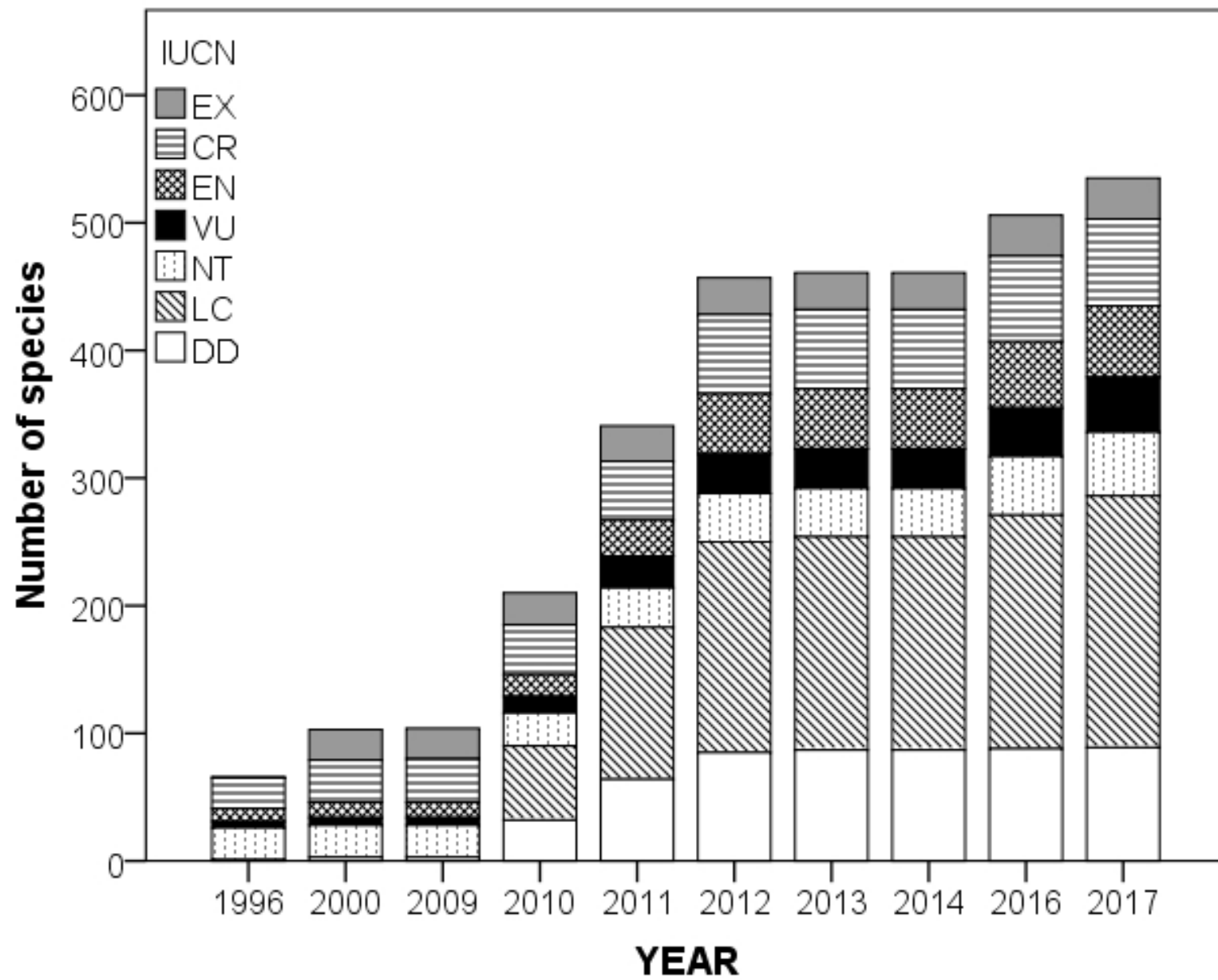
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861 **Table 1**

862 The top 20 selected research priorities for assessing freshwater mussel conservation status at the species level. Research
 863 priorities are grouped into intrinsic or extrinsic factors. Intrinsic factors are measures of species condition, and extrinsic
 864 factors describe the state of the environment.

Category	Factor	Research priority
Intrinsic	Species identity	Perform accurate species identification and determine species boundaries
	Population size and species distribution	Determine species' population size; and current species distribution
	Population trends	Determine the historic distribution range and population size
	Demography	Assess population viability, and perform demographic studies (e.g., recruitment, population age structure, mortality rates, immigration and emigration)
	Life-history traits	Identify traits and boundaries related to extinction risk, particularly characterize reproductive traits (reproductive strategy, timing of the reproductive cycle, fecundity, lifespan, age at sexual maturity, and generation length), and determine life history traits
Extrinsic	Fish hosts	Identify primary mussel/host relationships, host histological compatibility (metamorphosis success), and host availability (contact probability)
	Habitat characteristics	Identify habitat characteristics (including climate and catchment characteristics) related to species presence
	Catchment usage	Analyze species' response to land-use change (sedimentation, turbidity, nutrient pollution)
	Habitat alteration	Assess the impact of impoundments or other alterations on mussels (e.g., low oxygen concentration, increasing temperature, stagnation), and assess the impact of impoundments or other alterations on fish host movements and accessibility
	Water quality and sediment contamination	Assess ecotoxicological response of adult and early life stages (especially) to pollutants including novel contaminants
	Climate change	Determine the effect of increasing temperature at different organizational levels, determine the tolerance to emersion and assess the effect of successive extreme climatic events (e.g., heat waves, floods)
	Non-native bivalve species	Assess the strength of competition at different life stages solely or in combination with other threats (e.g., climate change, pollution)

Other extrinsic factors Identify and assess whether secondary factors (listed in text) represent a global or a localized threat to species



Extrinsic factors

Climate change

Catchment usage

Water habitat alteration

Intrinsic factors

Gamete production

Diseases parasitism

Fertilisation

Reproductive output

Invasive species

Infestation

Fish hosts

Recruitment

Predation

Population

Exploitation

Distribution

